

RESEARCH ARTICLE

Open Access



Ventilator-associated Pneumonia caused by commensal oropharyngeal Flora: a retrospective Analysis of a prospectively collected Database

Johannes B. J. Scholte^{1*}, Johan I. M. van der Velde², Catharina F. M. Linssen³, Helke A. van Dessel⁴, Dennis C. J. J. Bergmans², Paul H. M. Savelkoul², Paul M. H. J. Roekaerts⁴ and Walther N. K. A. van Mook²

Abstract

Background: The significance of commensal oropharyngeal flora (COF) as a potential cause of ventilator-associated pneumonia (VAP) is scarcely investigated and consequently unknown. Therefore, the aim of this study was to explore whether COF may cause VAP.

Methods: Retrospective clinical, microbiological and radiographic analysis of all prospectively collected suspected VAP cases in which bronchoalveolar lavage fluid exclusively yielded $\geq 10^4$ cfu/ml COF during a 9.5-year period. Characteristics of 899 recent intensive care unit (ICU) admissions were used as a reference population.

Results: Out of the prospectively collected database containing 159 VAP cases, 23 patients were included. In these patients, VAP developed after a median of 8 days of mechanical ventilation. The patients faced a prolonged total ICU length of stay (35 days [$P < .001$]), hospital length of stay (45 days [$P = .001$]), and a trend to higher mortality (39 % vs. 26 %, [$P = .158$]; standardized mortality ratio 1.26 vs. 0.77, [$P = .137$]) compared to the reference population. After clinical, microbiological and radiographic analysis, COF was the most likely cause of respiratory deterioration in 15 patients (9.4 % of all VAP cases) and a possible cause in 2 patients.

Conclusion: Commensal oropharyngeal flora appears to be a potential cause of VAP in limited numbers of ICU patients as is probably associated with an increased length of stay in both ICU and hospital. As COF-VAP develops late in the course of ICU admission, it is possibly associated with the immunocompromised status of ICU patients.

Keywords: Ventilator-associated pneumonia, Bronchoalveolar lavage, Commensal oropharyngeal flora, Intensive care unit, MALDI-TOF-MS

Background

Mechanically ventilated patients are at risk for ventilator-associated pneumonia (VAP) [1], which is associated with an increased intensive care unit (ICU) length of stay (LOS), morbidity and mortality [2–5]. A globally accepted gold standard for the diagnosis of VAP is lacking [6–8], with the exception of the new surveillance ventilator-associated events definitions [9]. Generally recommended and accepted microbiological diagnostic modalities for VAP are quantitative specimen collection techniques as bronchoalveolar lavage (BAL) [10, 11] and protected

specimen brushing obtained bronchoscopically and non-bronchoscopically. The identification of aerobic oropharyngeal flora in BAL fluid (BALF) cultures is commonly considered colonization, contamination from the upper respiratory tract or otherwise regarded clinically irrelevant, regardless of quantity [11–13].

Authoritative guidelines remain inconclusive regarding the role of commensal oropharyngeal flora (COF) as a causative agent in VAP, mainly due to a scarcity of studies in this research field [14]. However, there is evidence that COF may cause pulmonary infection, mostly in immunocompromised patients. Examples are *Streptococcus oralis/mitis* in neutropenic patients [15, 16] and species of the *Streptococcus anginosus* group in patients with predisposing factors including cancer,

* Correspondence: janscholte@hotmail.com

¹Department of Intensive Care Medicine, Luzerner Kantonspital, 6000 Luzern 16, Switzerland

Full list of author information is available at the end of the article

alcoholism [17, 18], and cystic fibrosis [19, 20]. Furthermore, aspiration may cause a pneumonia due to both anaerobic and/or aerobic oropharyngeal flora [21]. For the critically ill, several studies demonstrate that the innate immune response is declined due to several different mechanisms [22–24]. Therefore, one may hypothesize that ICU patients are likewise at risk for infections by COF. This study explores whether COF can be a cause of VAP.

Methods

Setting

The study was conducted at the Maastricht University Medical Centre, a 715-bed hospital with approximately 30,000 annual admissions, 18 mixed surgical-medical ICU beds, and 9 post-cardiothoracic surgery beds. Other elective postoperative patients are rarely admitted, due to a 24-h post anaesthesia care unit. When technically possible and safe, a BAL was performed in all mechanically ventilated patients who met the clinical criteria of suspected VAP. These criteria include (≥ 2 of the following) a rectal temperature > 38.0 °C or < 35.5 °C, white blood cell count $> 10,000/\mu\text{l}$ or $< 3,000/\mu\text{l}$, purulent sputum, and a new, persistent or progressive infiltrate on chest X-ray [25]. In patients with localized pulmonary lesions, the affected region was sampled, whereas in case of diffuse pulmonary lesions the middle lobe or lingula was lavaged. The BALF was microbiologically evaluated within 15 min after it was obtained. Selective oropharyngeal decontamination (SOD) is used since December 2010, whereas selective digestive tract decontamination (SDD) is used since January 2012. The SOD consists of topical antibiotics (polymyxin E, tobramycin, amphotericin B) applied to the oropharynx, whereas the SDD consists of oropharyngeal and gastric application of the same non-absorbable antibiotics along with a four day course of intravenous cefotaxime. The ethics committee of the institution, the “Medical Research Ethics Committee”, approved the study and informed consent was regarded unnecessary since standard care was provided.

Definitions

Ventilator-associated pneumonia in clinically suspected cases (for definition, see previous paragraph) was diagnosed if subsequent BALF analysis was indicative for pneumonia: cultures yielding a potentially pathogenic microorganism [12] $\geq 10^4$ cfu/ml and/or if ≥ 2 % BALF cells containing intracellular organisms (ICOs) [10, 26]. In a pneumonia suspected case that was admitted from home less than 3 days prior to diagnosis, community-acquired pneumonia (CAP) was considered, if this case had no recent contact with the healthcare system. A CAP was also considered when the potentially pathogenic microorganism was very unlikely to be nosocomial (e.g. *Haemophilus influenzae*, *Mycoplasma* spp.) [27].

When a clinical suspected case was admitted in the hospital for more than 3 days and with positive BALF results, but was not mechanically ventilated for ≥ 48 h in the 72 h prior to the pneumonia, hospital-acquired pneumonia (HAP) was diagnosed [14].

Commensal oropharyngeal flora as the cause of VAP was considered in VAP suspected cases if BALF quantitative cultures revealed COF $\geq 10^4$ cfu/ml without significant growth ($\geq 10^4$ cfu/ml) of other potentially pathogenic microorganisms. Commensal oropharyngeal flora included (a combination of) the following bacteria: viridans streptococci, coagulase-negative staphylococci, *Haemophilus* spp. (excluding *H. influenzae* if not predominant), *Moraxella* spp. (if not predominant), *Corynebacterium* spp., *Neisseria* spp., *Peptostreptococcus* spp., *Stomatococcus* spp., and *Prevotella* spp. [28]. Whereas *Candida* spp. may occasionally cause a pneumonia [29], *Candida* spp. were considered nonpathogenic in this study, consistent with previous studies and guidelines [30–32].

Data collection

From January 2005 until January 2014, all results of BALF analyses from patients consecutively admitted to the ICU were prospectively collected. From this database, patients with suspected COF as the cause for VAP were included in the present study. Cases lacking a microbiological BAL report were excluded. Retrospectively, the following clinical data were collected or calculated from the included cases: body temperature, C-reactive protein (CRP), white blood cell count, antibiotic administration, ICU length of stay, hospital length of stay, duration of mechanical ventilation, mortality, acute physiology and chronic health evaluation (APACHE)-II score (to calculate standardized mortality ratio [SMR; observed mortality divided by expected mortality]) [33], sequential organ failure assessment (SOFA) score (to determine the extent of critical illness) [34, 35], clinical pulmonary infection score (CPIS; more than 6 points is indicative of pneumonia) [36], and post-mortem examination, if available.

Reference population

On the advice of the statistical department a reference population was used in order to place the results of suspected cases in perspective. Since June 2013, the hospital participated in the Dutch National Intensive Care Evaluation registry. To experience the possibilities of this registry, the characteristics of all patients admitted from June 2013 to April 2014 in the same ICU were extracted from this database. Post cardiothoracic surgery patients (44 % of all ICU admissions) were excluded. It should be realized that a reference population is not a control

group and interpretation of finding should be performed in this perspective.

Microbiological data collection

Bronchoalveolar lavage fluids were initially analysed according to a highly standardized protocol as described elsewhere [37]. From each BALF sample, 6 ml was centrifuged (250 g for 10 min), dividing the sample into cells and supernatant. The supernatant was stored in tubes of 1 ml at -80°C . The cells were re-suspended in 6 ml of a mixture of Eagle's Minimal Essential Medium with 2 % Dimethyl Sulfoxide and stored in tubes of 1 ml at -80°C . Oropharyngeal flora was (formerly) reported on the basis of classical bacteriological phenotypic identification tests. In order to confirm and specify these results, included samples were defrosted and quantitatively cultured on blood agar, chocolate agar, and MacConkey agar. The different colony types were identified using matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF-MS), a rapid and highly accurate soft ionization technique [38]. Antibiotic susceptibility was assessed for all separate strains when quantitative cultures revealed $\geq 10^4$ cfu/ml. Results of polymerase chain reactions (PCRs) for viruses and *Pneumocystis jirovecii*, as well as Grocott's methenamine silver staining, were collected from electronic patient data organisers using SAP. In patients that were both admitted to the hospital less than 8 days prior to BAL procedure and that lacked PCR results, PCRs for the identification of respiratory viruses were performed on the defrosted BALF. These viruses included influenza virus A and B, human respiratory syncytial virus, human metapneumovirus and parainfluenza virus 1–4. Furthermore, results of endotracheal aspirates (ETA) were analysed, preferably from the day of BAL, otherwise one day before or after. Endotracheal aspirates were obtained twice weekly and in case of clinical suspicion of a pulmonary infection. Samples were immediately microbiologically evaluated when obtained during daytime.

Commensal oropharyngeal flora caused ventilator-associated pneumonia likelihood

Based on the collected clinical, radiographic, and microbiological data and subsequent microbiological analyses made, the likelihood of presence of COF caused VAP (COF-VAP) was evaluated by 4 researchers (two consultant ICU physicians, one medical microbiologist, one ICU researcher).

Statistics

Patient characteristics were analysed using descriptive statistics and presented as the mean \pm standard deviation, median including interquartile range, or absolute numbers and percentages of patients, where applicable.

Demographic and clinical characteristics of the suspected cases were compared with the reference population using the single sample *t*-test, the paired samples *t*-test or the Fisher's exact test, where appropriate. For the comparison of the observed and expected mortality, a Chi-square test was used. A one-way within-subjects ANOVA was used to analyse the course of the SOFA-score, body temperature, leukocytes and CRP. Statistical significance was defined as $P < .05$. The IBM SPSS Statistics version 20 for Windows (Chicago, IL, USA) was used for analysis.

Results

The study was conducted between January 2005 and July 2014. During this period, 17,254 patients were admitted to the ICU, of which 159 individuals met the VAP criteria during the study period. Twenty-three patients were considered to have COF-VAP (see Fig. 1 for inclusion flow chart). During the defined period, 899 patients (excluding post cardiothoracic surgery patients) needed ICU admission and consequently, these patients were included in the reference population.

Basic characteristics including clinical findings, BALF cytological results and outcome of the studied group and the reference population are presented in Table 1. Individual results of the analyses are presented in Table 2. The most common reason for ICU admission was respiratory failure (7/23 [30 %]) followed by multi-trauma (5/23 [22 %]). The reason for ICU admission of the study group was more often surgically related compared to the reference population (13/23 [57 %] vs. 267/899 [30 %], $P = .01$). Suspected COF-VAP developed after a

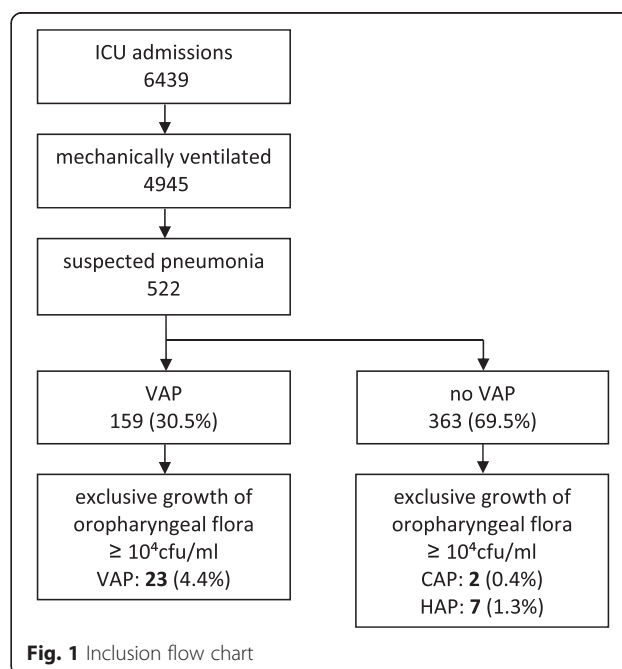


Table 1 Basic characteristics and clinical findings

	Commensal oropharyngeal flora VAP N = 23	Reference population N = 899	P value
Age, mean (SD)	55 (16)		
Male (%)	16 (70 %)	557 (62 %)	.52
Medical history (%)			
Cardiac	12 (52 %)		
Pulmonary	7 (30 %)		
Oncologic	7 (30 %)		
Immunocompromised	2 (9 %)	98 (11 %)	.85
Indication for ICU admission (%)			
Surgical	13 (57 %)	267 (30 %)	.01
Non-surgical	10 (43 %)	632 (70 %)	.01
ICU admission scores (SD) n = 21			
APACHE-II score	19.8 (9.9)	19.1 (8.4)	.75
Predicted mortality	31 % (26)	34 % (26)	.60
SOFA score	8.1 (3.3)		
Characteristics at the day of diagnosis (SD)			
SOFA score	6.9 (3.7)		
Body temperature (°C)	38.3 (1.1)		
WBC count (10 ⁹ /L)	14.3 (8.4)		
CRP (mg/L)	180 (108)		
CPIS	6.4 (2.1)		
Median days on MV (IQR)	8 (4–19)		
Number of BAL fluid cells	6.9*10 ⁵ (1.7*10 ⁵)		
of which PMNs	60 % (35)		
of which containing ICOs	5.1 (6.5)		
Outcome			
ICU mortality	9 (39 %)	234 (26 %)	.16
SMR (SD)	1.26 (0.42)	0.77 (0.04)	.14
28-day mortality	3 (13 %)		
60-day mortality	6 (26 %)		
Median hospital LOS (IQR)	63 (50–146)	23.7 (=mean)	.001
Median ICU LOS (IQR)	33 (13–62)	5.7 (=mean)	.000
Median days on MV (IQR)	34 (17–67)		
Median hospital LOS after diagnosis (IQR)	22 (10–59)		
Median ICU LOS after diagnosis (IQR)	13 (7–55)		

If no number is provided, this information was not available in the Netherlands intensive care evaluation registry. APACHE-II score could not be calculated in 2 patients due to incomplete admission data

median of 8 days of mechanical ventilation. Aspiration prior to BAL was considered possible in 3 patients. Five patients were mechanically ventilated for more than 2 days by means of a tracheostomy prior to the suspected VAP.

Clinical parameters

On the day of BAL, mean CPIS revealed 6.4 (standard deviation 2.4). An increase in mean SOFA score, white blood cell count, and CRP was observed on the day after the diagnosis compared to the day of diagnosis (See Fig. 2). After this initial increase, a gradual and persistent decrease was observed for all parameters.

Outcome

As shown in Table 1, median hospital LOS and ICU LOS in the studied group were significantly longer compared to the reference population (64 days vs. 24 days [$P = .001$] and 33 vs. 5.7 days [$P < .001$], respectively). The SMR in the studied group revealed 1.26 compared to 0.77 in the reference population ($P = .137$).

Microbiological results

Overall cytological BAL findings are presented in Table 1 and an individual overview of cytological and microbiological analysis is provided in Table 2. In 21 patients, a BALF cell count was performed revealing 13 BALF with more than 50 % polymorphonuclear neutrophils. In 11 BALF, 2 % or more BALF cells contained ICOs. Additionally, the percentage of squamous epithelial cells, an indicator of oropharyngeal contamination, was less than 1 % in all patients. In 6 patients, BALF could not be retrieved from storage and in 3 patients, BALF culture yielded no growth. These 3 fluid samples were preserved for 5, 6, and 9 years. In the remaining 14 BALF, MALDI-TOF-MS was able to identify 1 or more bacterial species. In 5 patients, bacteria $\geq 10^4$ cfu/ml were identified, of which 2 types are potential pathogenic pathogens (*Citrobacter koseri* 10^6 cfu/ml and *H. influenzae* 10^6 cfu/ml) and 1 type possibly pathogenic (*E. faecalis* 2×10^4 cfu/ml). In 4 BALF, the presence of exclusively COF according to the definition was confirmed by MALDI-TOF-MS.

Endotracheal aspirates were obtained from all studied cases. Rare, moderate and heavy growth of COF was present in 9, 1, and 4 cases, respectively. Moderate or heavy growth of COF was present in 2 out of 15 patients with probably COF-VAP [sensitivity 13 %]. In 6 cases, moderate or heavy growth of another pathogenic microorganism was identified (*S. aureus* [twice], *P. aeruginosa* [twice], *H. influenzae* [case described above in this section], and *M. morgani*). Three cases were eventually diagnosed with pneumonia caused by the same microorganism (positive predictive value 50 %). PCR on BALF for viral pathogens was performed in 15 patients.

Table 2 Characteristics, diagnostic results and outcome in suspected cases of commensal oropharyngeal flora caused ventilator-associated pneumonia

Case	Sex	Admission indication	BAL fluid analysis			Endotracheal aspirate Semiquantitatively	Antibiotic treatment (days [0 = diagnose])	Additional characteristics	Diagnosis	ICU LOS
			% ICOs	culture results in cfu/ml	MALDI-TOF -MS and re-culturing (cfu/ml)					
1	M	Respiratory failure	0.0	COF 6*10 ⁴	No growth	COF few	Co-trimoxazole -7-27	Some COF possibly resistant to antibiotics	COF-VAP	92 ^a
			60	11.6			Piperacillin -7-27			
2	F	Abdominal sepsis	0.0	COF 5*10 ⁴	Specimen storage lacking	<i>P. aeruginosa</i> heavy <i>C. albicans</i> few	Piper/tazob -4-11	Tracheostomy	COF-VAP or <i>P. aeruginosa</i> VAP	31
			77	87.2	<i>P. aeruginosa</i> 2*10 ³ , sensitive to piper/tazob					
3	F	Abdominal sepsis	8.2	COF 2*10 ⁵	<i>R. dentocariosa</i> 2*10 ²	<i>S. aureus</i> heavy	Piper/tazob 0-2	Nocturnal CPAP for OSAS	COF-VAP	60
			57	85.6	<i>S. oralis</i> 7*10 ² <i>S. aureus</i> 6*10 ²		Vancomycin 2-9			
4	F	Post cardiac arrest	10.8	COF 10 ⁵	<i>S. constellatus</i> 10 ⁶	<i>S. milleri</i> heavy	Piper/tazob 0-3	Possible aspiration	COF-VAP	33
			67	34.6	<i>S. milleri</i> 10 ⁵ <i>S. epidermidis</i> 3*10 ³		Penicillin 3-8			
5	M	Multi-trauma	2.5	COF 4*10 ⁴	<i>S. mitis/oralis</i> 5*10 ³	No growth	Amoxi/clav acid 1-8	Chronic obstructive pulmonary disease	COF-VAP	16
			47	91.5	<i>N. mucosa</i> 10 ³ <i>C. sputigena</i> 5*10 ²					
6	M	Neurological	4.8	COF 10 ⁵	<i>S. mitis/oralis</i> 5*10 ³	<i>Citrobacter</i> spp. few	Piper/tazob 0-6		COF-VAP	15
			23	68.2	<i>S. aureus</i> 10 ³ <i>E. cloacae</i> 10 ³					
7	M	Multi-trauma	8.8	<i>P. melaninogenetica</i> 10 ⁵	Specimen storage lacking	COF few	Metronidazole 1-7	<i>P. melaninogenetica</i> not susceptible to Metronidazole	COF-VAP	9
			39	90.8						
8	M	Pneumococcal pneumonia	2.0	COF 10 ⁵	Specimen storage lacking	<i>P. aeruginosa</i> moderate	Ciprofloxacin 0-13	Tracheostomy. Pulmonary rehabilitation clinic.	COF-VAP	134
			64	21.2	<i>P. aeruginosa</i> 2*10 ²					

Table 2 Characteristics, diagnostic results and outcome in suspected cases of commensal oropharyngeal flora caused ventilator-associated pneumonia (Continued)

9	M	Respiratory failure	1.4	COF 3*10 ⁴	Specimen storage lacking	COF few	Gentamicin 0	Tracheostomy	COF-VAP	47 ^a
			67	23.6			Ciprofloxacin -5-8 Metronidazole -5-8 Vancomycin -5-8	Post-mortem exam: severe chronic fibrotic inflammation and active pneumonia		
10	M	Respiratory failure	0.2	COF 10 ⁵	No growth	<i>C. albicans</i> rare	Ciprofloxacin -14-0	Lobectomy for aspergilloma. Impossible to wean from MV.	COF-VAP	70 ^a
			62	99.6		<i>C. albicans</i> 10 ⁵	Piper/tazob -2-3 Fluconazole 3-17			
11	M	Cerebral hemorrhage	16	COF 10 ⁴	<i>L. acidophilus</i> 10 ²	<i>H. influenzae</i> few	Piper/tazob 0-2		COF-VAP	11
			31	95.4		<i>P. melaninogenetica</i> 2*10 ²	Amoxi/clav acid 2-8			
12	M	Sepsis	0.0	COF >10 ⁵	<i>M. morgani</i> 2*10 ³	<i>M. morgani</i> moderate	Flucloxacilline -22- -3		COF-VAP	42 ^a
			80	15.4	<i>S. malthophilia</i> 5*10 ²	<i>C. albicans</i> moderate <i>K. pneumoniae</i> few	Rifampicine -17- -10 Piper/tazob -3 -10 Co-trimoxazole 3-10			
13	M	Post cardiothoracic surgery	19.6	COF 3*10 ⁴	<i>S. anginosus</i> 10 ³	COF moderate	Piper/tazob 0-8			
		COF-VAP	63 ^a							
			71	56.2	<i>S. constellatus</i> 10 ³	<i>C. freundii</i> moderate				
					<i>N. mucosa</i> 10 ²					
14	M	Multi-trauma	3.4	COF 10 ⁵	<i>E. faecalis</i> 6*10 ²	COF heavy	Piper/tazob 0-1	Tracheostomy	COF-VAP	62
			41	44.4	<i>P. aeruginosa</i> 7*10 ³	<i>S. mitis/oralis</i> 5*10 ²	<i>P. aeruginosa</i> heavy Amoxi/clav acid 1-7			
15	M	Respiratory failure	NP	COF 10 ⁴	<i>E. faecalis</i> 2*10 ⁴	COF few	Co-trimoxazole -25 -13	Aids. ADV ct 17. CMV ct 37. <i>P. jirovecii</i> +. CT-thorax suggestive for PcP.	PcP CAP, ADV CAP, and/or <i>E. faecalis</i> VAP	44 ^a
			40	NP	<i>S. hominis</i> 2*10 ²					

Table 2 Characteristics, diagnostic results and outcome in suspected cases of commensal oropharyngeal flora caused ventilator-associated pneumonia (Continued)

							Piper/tazob 0-7	Post-mortem exam: active pneumonia, possibly PcP, ADV and CMV.		
							Vancomycin 0-8			
16	F	Respiratory failure	0.0	COF 10 ⁵	No growth	COF few	Piper/tazob -14- -4	Ileus. Post-mortem exam: faecal peritonitis. No pneumonia. Culture -	Abdominal sepsis	4 ^a
	55		78.0	<i>C. glabrata</i> 4*10 ²		<i>C. albicans</i>	Ciprofloxacin -3-1			
				<i>C. albicans</i> 2*10 ²			Vancomycin -2- 2			
17	F	Post abdominal surgery	0.0	COF 3*10 ⁴	Specimen storage lacking	COF few	Missing data		COF-VAP	13
	70		68.0							
18	M	Post neurological surgery	1.6	COF 2*10 ⁴	<i>S. aureus</i> 3*10 ³	<i>S. aureus</i> heavy	Piper/tazob ?-0		COF-VAP or <i>S. aureus</i> VAP	17
	58		94.4	<i>S. aureus</i> 3*10 ³ , sensitive to piper/tazob			Flucloxacillin 0- 10			
							Gentamicin 3-5			
19	M	Post- cardiothoracic surgery	0.0	COF 5*10 ⁴	Specimen storage lacking	COF few	Piper/tazob -8 - -1	Tracheostomy	COF-VAP	81
	73		2.8			<i>K. pneumoniae</i>	Meropenem -1- 7			
20	M	Multi-trauma	19.4	COF 10 ⁴	<i>H. influenza</i> 10 ⁶	<i>H. influenza</i> heavy	Amoxi/clav acid -5- -4	Possible aspiration	<i>H. influenza</i> CAP	62
	49		95.0		<i>E. coli</i> 4*10 ³	OF few	Piper/tazob 0-5			
					<i>N. subflava</i> 4*10 ³	<i>S. aureus</i> few	Gentamicin 4			
21	F	Post cardiac arrest	0.0	COF 10 ⁴	<i>E. faecium</i> 5*10 ³	COF heavy	Piper/tazob 0-3	B- cel CLL. PIV-3 ct 19.	PIV-3 CAP or <i>E. faecium</i> VAP	22 ^a
	45		1.0		<i>E. faecalis</i> 9*10 ²		Vancomycin 0-3	Post-mortem exam: active pneumonia.		
					<i>S. epidermidis</i> 6*10 ²		Meropenem 2-3	Cultures: CNS, <i>E. faecium</i> and <i>Candida</i> spp.		

Table 2 Characteristics, diagnostic results and outcome in suspected cases of commensal oropharyngeal flora caused ventilator-associated pneumonia (Continued)

22	F	Cerebral hemorrhage	7.4	COF 10 ⁴	<i>C. koseri</i> 10 ⁶	COF heavy	Vancomycin 0-2	<i>C. koseri</i> resistant to vancomycin. Possible aspiration.	<i>C. koseri</i> VAP	3 ^a
			58	85.6	<i>C. glabrata</i> 10 ³					
23	M	Respiratory failure	NP	COF 10 ⁴	<i>S. mitis/oralis</i> 10 ⁴	COF few	Amoxi/clav acid -18- -2	Admitted with bilateral pneumonia. <i>M. pneumoniae</i> on day 7 (ct 23. IgG +)	<i>M. pneumoniae</i> CAP	11
			25	NP	<i>S. haemolyticus</i> 10 ³		Ciprofloxacin -8- -2			
					<i>N. subflava</i> 6*10 ²		Azitromycin 0-4			

^adeceased

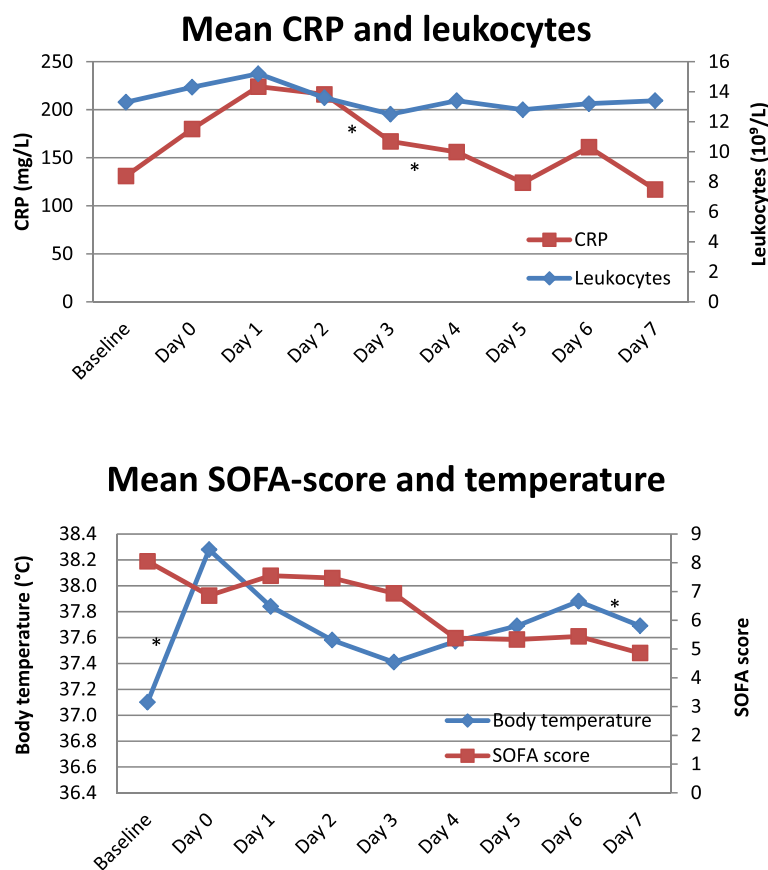


Fig. 2 Course of clinical and laboratory parameters. Numbers signify mean values in all commensal oropharyngeal flora caused ventilator-associated pneumonia suspected cases. *P value < .05 for the increase/decrease of the value as compared to the previous value

Parainfluenza virus-3 (cycle threshold (ct) 19 [infection likely]) and Cytomegalovirus (ct 37 [probably reactivation]) were identified, each virus in 1 case. In 1 patient, *Mycoplasma pneumoniae* (ct 23 [infection likely]) was identified by PCR. In 1 case, both BALF Grocott's staining and PCR were positive for *Pneumocystis jirovecii*.

Antibiotic treatment

Data concerning the administered antibiotics were available in 22 patients. Eleven of these patients (50 %) received (broad spectrum) antibiotics in the 72 h prior to the BAL. Overall, piperacillin/tazobactam was the most frequently used antibiotic (15/22 [68 %]). Five patients received inadequate treatment for COF-VAP (See Table 2; Patients no. 7, 8, 10, 22, and 23) of which 2 had unfavourable respiratory outcome (patients no. 8 and 10) and 2 died (patients no. 10 and 22), 47 and 3 days after the diagnosis, respectively. Of the 17 patients that received appropriate COF-VAP treatment, 6 patients died after a median of 54 days after the diagnosis.

Post mortem examination

In 4 out of the 9 deceased patients, consent for autopsy was obtained. Pathological evidence of an active pneumonia was found in 3 patients in post-mortem examination, performed at least 22 days after diagnosis. A faecal peritonitis was observed in the fourth patient, probably explaining her respiratory deterioration.

Level of certainty of the presence of commensal oropharyngeal flora ventilator-associated pneumonia

An alternative diagnosis was concerned more likely in 6 patients: *Citrobacter koseri* VAP, *H. influenzae* pneumonia, parainfluenza virus-3 CAP or *E. faecium* HAP, *Mycoplasma pneumoniae* CAP (all 4 with clinical signs of pneumonia), *Pneumocystis jirovecii* caused CAP (with CT-thorax suggestive of *Pneumocystis* pneumonia), and abdominal sepsis (with clinical signs of an ileus the day before the patient deceased). Eleven patients received antibiotics in the three days prior to the (presumed) VAP, making their culture results more difficult to

interpret. The cultures in cases number 2 and 18 yielded $\geq 10^3$ cfu/ml *P. aeruginosa* and *S. aureus*, respectively, under appropriate antibiotic treatment, which cannot exclude their causative contribution to VAP. Therefore these 2 cases were considered to have possible COF-VAP. In the remaining 15 cases (9.4 % of all VAP cases), no other infection was found and COF-VAP was considered likely.

Additional findings

Additionally, 3 cases of probable COF-HAP and 1 case of probable COF-HAP were identified in 8 eligible cases (see Appendix). Besides the fact that these 4 positive cases were immunocompromised due to previous used medication or malignancies, further discussion of this subgroup is beyond the scope of this article.

Discussion

This large retrospective analysis of a prospectively collected case series revealed 23 patients suspected of COF-VAP. In 15 patients (9.4 % of all VAP cases), COF-VAP was the most likely diagnosis.

Previous studies

So far, 1 currently available study investigated the possible role of COF as the cause of VAP [39]. This French group retrospectively investigated 29 episodes of VAP with significant growth of oropharyngeal and cutaneous commensal microorganisms in BALF and protected brush specimen derived from 28 immunocompetent patients. The suspected cases also revealed signs of VAP including a longer ICU LOS. A panel of 3 experts confirmed 14 out of the 29 (48 %) suspected cases as COF-VAP (corresponding to 3.8 % of their VAP cases), whereas in the other cases an alternative diagnosis was considered more likely. The authors concluded that COF may cause VAP in ICU patients with no known prior immunodeficiency, that it has similar clinical features as VAP, and that patients should be treated with antibiotics with the probable exception of stable patients with a high likelihood of an alternative diagnosis [39]. The results of the present study, using more clinical data and microbiological analyses, are in support of their preliminary findings.

Two decades ago, several small studies revealed the presence of COF in post-mortem lung biopsy cultures in previously mechanically ventilated patients [40–42] (reported incidences varied from 9 % to 57 %, with not all studies reporting the level of growth), whereas another study found no COF in post-mortem biopsy diagnosed VAP cases [43].

Epidemiology

One might perhaps expect a rise in COF-VAP rates after the introduction of SOD/SDD, as it focusses on the elimination of Gram negatives as well as the selection of commensal bacteria [44]. Yet, so far (see Table 2) no diagnosis of COF-VAP was made after the introduction of SOD/SDD, whereas the incidence of VAP per 1,000 ventilator days declined from 4.38 ± 1.64 before to 1.64 ± 0.43 ($P = .007$) after the introduction of SOD and SDD in our clinic [45].

Results of endotracheal aspirates

Whereas ETA and BALF analysis can both be used to diagnose VAP [14, 30], results of these 2 diagnostic modalities agree only fairly [46]. Overall, the current results confirm the poor agreement between BALF and ETA analyses results.

Re-cultured bronchoalveolar lavage fluid and identification with matrix-assisted laser desorption/ionization time-of-flight mass spectrometry

Whereas all 23 BALF samples revealed $\geq 10^4$ cfu/ml COF at the time of the BAL procedure, the repeated microbiological examinations on preserved specimen were frequently not in accordance. The BALF samples that revealed no growth were similar durations as the samples that did reveal growth, suggesting that the time of storage did not cause the disagreement. However, *Neisseria* spp., *Streptococcus* spp. (not *S. pyogenes*), and *Haemophilus* spp. are known with a limited survival on inanimate surfaces [47], contributing to the absence of growth on re-culturing despite the correct solvents and temperature. The lack of significant growth of microorganism could have been related to the storage-related factors, as well as the presence of antibiotics in the sample before storage. On the other hand, one may doubt if the *C. koserii*, *H. influenzae*, and *E. faecalis* identified by MALDI-TOF-MS were true pathogens or that their presence was caused by contamination before, during, or after the BALF storage and retrieval.

Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry was able to identify all strains that re-grew on any agar plate. *Rothia dentocariosa*, *Capnocytophaga sputigena*, and *Lactobacillus acidophilus* are part of COF, although not included in our definition since these species were not routinely identified before MALDI-TOF-MS use.

Diagnosis

Although some results are more difficult to interpret due to previous antibiotic use, identified COF is mostly susceptible to the antibiotics used, but was nonetheless still present in significant numbers in BALF. Furthermore, at least 9 patients that were eventually diagnosed

with COF-VAP did not receive antibiotics prior to BAL. In 6 patients, an alternative diagnosis was concerned more likely, although COF as a contributing or the main cause of VAP can neither be established nor excluded with certainty. Remarkably, 3 suspected VAP cases actually had CAP.

Treatment and outcome

Four out of the 15 patients with probable COF-VAP received inappropriate antibiotic treatment. The respiratory outcome was unfavourable in 2 out of 3 cases that survived the VAP episode, suggesting that COF-VAP should be treated appropriately. Commensal oropharyngeal flora is generally susceptible to the antibiotics suggested by authoritative guidelines [14].

Entity or fiction?

Although no hard argument can be provided to state that COF-VAP is an entity, 6 arguments originated from the present study support that COF indeed may cause VAP. First, it is plausible from a pathophysiological point of view. As stated in the introduction section, COF may behave pathogenic in immunocompromised patients. Whereas COF-VAP developed after a median of 8 days of mechanical ventilation, a previously immunocompetent person may already be considered immunocompromised after 48 h of ICU admission [24]. Additionally, many COF-VAP suspected cases revealed positive viral and fungal PCR, which is also associated with a decreased immune status. Finally, COF was able to cause HAP and CAP in immunocompromised patients. Second, scores resulting from generally accepted clinical scoring systems, as well as laboratory results, indicated a significant clinical relevance. Third, cytological analysis of BALF frequently was indicative of a bacterial infection. Fourth, COF was the most likely cause of VAP in 15 cases and possibly the cause in 2 cases. Fifth, inappropriate antibiotic treatment for COF-VAP ($n = 4$) was associated with unfavourable respiratory outcome. Sixth and last, COF-VAP was associated with increased ICU and hospital LOS including a trend towards increased mortality as compared to a reference population.

Contrariwise, post-mortem examination 4 days after BAL of one patient revealed no signs of pneumonia, indicative of a false positive BALF analysis. However, at the time of BAL, this patient presents with an ileus and abdominal sepsis was the post-mortem diagnosis. Although it is possible that the patient fulfilled all criteria for performing a BAL due to the extrapulmonary problems, BAL should thus only be performed in the absence of an obvious alternative explanation for the patient's clinical presentation.

The current study demonstrated a probable association between merely significant growth of COF in VAP

suspected patients and worse outcome. Similar to VAP caused by other microorganisms [2], it remains unknown whether this association is causal or that COF-VAP results from critical illness and that the outcome is therefore unfavourable.

Limitations of the study

There are several limitations to the study. First, due to the lack of a globally accepted gold standard to diagnose VAP [7, 8, 11], VAP in clinically suspected patients was defined as BALF revealing ≥ 2 % ICOs and/or significant growth ($\geq 10^4$ cfu/ml [36]) of a potential pathogenic microorganism. Second, a limited number of clinical and microbiological data were irretrievable in a number of patients. Third, median ICU and hospital LOS of the suspected cases were compared to mean LOS in a recent patient population. Since mean ICU and hospital LOS declined during the studied period (2005–2013), this comparison may be unfair. However, since the mean ICU LOS was 8.6 days in 2005, a median ICU LOS of 33 days in the studied period is still significantly higher. Fourth, enterococci were excluded as a potential cause for VAP, although these microorganisms may possibly also cause VAP in the immunocompromised ICU-patient. Fifth, a reference population is not a control group. In future studies, a control group should preferably consist of “regular” VAP cases to demonstrate that COF behaves like “regular” VAP causative microorganisms. Finally, in the process of BALF storage and reculturing the occurrence of contamination cannot be ruled out.

Future studies

Since COF-VAP has a low overall incidence (15 out of 6500 ICU admissions [0.23 %]) and VAP incidences decline [45, 48], future research should preferably focus on multi-centre trials. Notwithstanding, when more similar studies become available, a meta-analysis could empower the results of Lambotte *et al.* and the current study.

Conclusions

As COF was identified as the most likely causative agent in 9.4 % of all VAP episodes, COF is probably an overlooked cause of VAP. The immunocompromised status of the ICU patient may contribute to its origination. Commensal oropharyngeal flora VAP is probably associated with significant clinical signs of bacterial infection, a prolonged ICU and hospital LOS, and a trend towards increased mortality. In the absence of other pulmonary and non-pulmonary explanations for a patient's pulmonary deterioration, ICU physicians should perhaps acknowledge this entity and treat it accordingly.

Appendix

Table 3 Individual characteristics, diagnostic results and outcome of the suspected cases of commensal oropharyngeal flora caused hospital-acquired pneumonia and community-acquired pneumonia

Case	Sex	Admission indication	BAL fluid analysis			Endotracheal aspirate Semiquantitatively	Antibiotic treatment (days [0 = diagnose])	Additional characteristics	Diagnosis	ICU LOS
			% ICO	BAL fluid culture cfu/ml	MALDI-TOF-MS and re-culturing (cfu/ml)					
Suspected cases of commensal oropharyngeal flora caused hospital-acquired pneumonia										
24	M	Respiratory failure	37.4	COF 10 ⁵	Specimen storage lacking	COF heavy	Piper/tazob -6 -1	Multiple myeloma. Post-mortem exam: CMV pneumonia. <i>Candida</i> spp.	CMV pneumonia	1 ^a
			65	33.0						
25	M	Respiratory failure	0.0	COF 10 ⁵	No growth	E. coli moderate	None	Immunosuppressive drug after lung transplantation. Ventilator depended after discharge	COF-HAP	182
			41	60.0						
26	F	Neurological	4.8	COF > 10 ⁵	<i>N. subflava</i> 4*10 ² <i>S. anginosus</i> 3*10 ² <i>S. mitis/oralis</i> 3*10 ² <i>H. influenzae</i> 2*10 ²	COF few	Amoxi/clav acid -2 -5	Possible aspiration Post-mortem exam: Indicative for amiodarone induced pneumonitis (AIP)	AIP	10 ^a
			73	63.8						
27	M	Hypokalemia	0.0	COF 10 ⁵	<i>E. faecium</i> 4*10 ⁴ <i>S. epidermidis</i> 3*10 ⁴	No growth	Ciprofloxacin -2 -2 Piper/tazob -2-18 Vancomycin 1-26	Acute myeloid leukemia. RSV ct 20, HSV-1 ct 29	RSV CAP/ <i>E. faecium</i> HAP	29 ^a
			60	96.8						
28	M	Respiratory failure	0.0	COF 10 ⁴	<i>C. jeikeium</i> 5*10 ³ <i>E. faecium</i> 4*10 ³	Candida spp.	Piper/tazob -11 -0 Meropenem 0-16	B-cel Lymphoma	COF-HAP	16 ^a
			50	11						
29	M	Respiratory failure	0.0	COF 10 ⁵	<i>E. faecium</i> 2*10 ⁴	COF few	Piper/tazob -2 -15 Vancomycin 4-12	Acute myeloid leukemia Possible aspiration. HSV-1 ct 34	COF-HAP	31 ^a
			54	0.8						
Suspected cases of commensal oropharyngeal flora caused community-acquired pneumonia										
30	F	Sepsis	0.0	COF 10 ⁴	No growth	NP	Amoxi/clavc acid 0-1	Metastatic mammacarcinoma	COF-CAP	1 ^a

Table 3 Individual characteristics, diagnostic results and outcome of the suspected cases of commensal oropharyngeal flora caused hospital-acquired pneumonia and community-acquired pneumonia (Continued)

	48		21.4				Ciprofloxacin 0-1			
31	M	Respiratory failure	0.2	COF 10 ⁴	<i>A. viscosus</i>	COF rare	Co-trimoxazole –20–27	Idiopathic CD4 deficiency. ICU admission for BAL only. <i>P. jiroveci</i> ct 24.	PcP	0
	68		32.8		<i>R. mucilaginosa</i> <i>S. mutans</i>					

^adeceased

Abbreviations, not used in the main text: HSV: Herpes simplex virus; RSV: respiratory syncytial virus

Abbreviations

ADV: Adenovirus; APACHE: Acute physiology and chronic health evaluation; BAL: Bronchoalveolar lavage; BAL: Bronchoalveolar lavage fluid; CAP: Community-acquired pneumonia; CLL: Chronic lymphocytic leukaemia; CMV: Cytomegalovirus; CNS: Coagulase-negative staphylococcus; CPAP: Continuous positive airway pressure; CPIS: Clinical pulmonary infection score; CRP: C-reactive protein; ct: Cycle threshold; ETA: Endotracheal aspirate; F: Female; HAP: Hospital-acquired pneumonia; ICO: Intracellular organism, in Table, percentage of BAL fluid cells containing intracellular organisms; ICU: Intensive care unit; IQR: Interquartile range; LOS: Length of stay; M: Male; MALDI-TOF-MS: Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry; MV: Mechanical ventilation; NP: Not performed; OSAS: Obstructive sleep apnoea syndrome; Pcp: *Pneumocystis pneumonia*; PCR: Polymerase chain reaction; PIV: Parainfluenza virus; PMN: Polymorphonuclear neutrophils; SD: Standard deviation; SDD: Selective digestive tract decontamination; SMR: Standardized mortality ratio; SOD: Selective oropharyngeal decontamination; SOFA: Sequential organ failure assessment; VAP: Ventilator-associated pneumonia.

Competing interests

The authors declare that they have no competing interest.

Authors' contributions

CFML initiated the study together with WNKAvM. JBJS, JIMvdV, and CFML prospectively collected and managed the database. JBJS and JIMvdV performed all statistical and initial interpretation of the results, wrote the first drafts of the manuscript and coordinated the implementation of suggestions from the other authors. The COF-VAP likelihood was evaluated by JBJS, JIMvdV, CFML, and WNKAvM. All authors actively participated in discussions on previous drafts and provided multiple qualitative comments resulting in the final approved manuscript.

Authors' information

JBJS, DCJJB and WNKAvM are all consultants in internal medicine and intensive care medicine. WNKAvM is also an associate professor in medical education and chair of the professional behaviour committee at Maastricht University. JIMvdV is a resident in anaesthesiology. CFML and HAvD are consultants in medical microbiology. PHMS is professor and chair of the department of medical microbiology and PMHJR is professor and chair of the department of intensive care medicine.

Acknowledgments

The advice and support in the statistical analysis by professor R. Vos of the department of statistics and methodology at Maastricht University are very much appreciated.

Author details

¹Department of Intensive Care Medicine, Luzerner Kantonspital, 6000 Luzern 16, Switzerland. ²Department of Intensive Care Medicine, Maastricht University Medical Centre+, P.O. box 5800, 6202 AZ Maastricht, The Netherlands. ³Department of Medical Microbiology, Atrium Medical Centre, P.O. box 4446, 6401 CX Heerlen, The Netherlands. ⁴Department of Medical Microbiology, Maastricht University Medical Centre+, P.O. box 5800, 6202 AZ Maastricht, The Netherlands.

Received: 7 March 2015 Accepted: 30 July 2015

Published online: 12 August 2015

References

- Klompas M, Khan Y, Kleinman K, Evans RS, Lloyd JF, Stevenson K, et al. Multicenter evaluation of a novel surveillance paradigm for complications of mechanical ventilation. *PLoS One*. 2011;6(3): e18062.
- Safdar N, Dezfulian C, Collard HR, Saint S. Clinical and economic consequences of ventilator-associated pneumonia: a systematic review. *Crit Care Med*. 2005;33(10):2184–93.
- Vincent JL, Rello J, Marshall J, Silva E, Anzueto A, Martin CD, et al. International study of the prevalence and outcomes of infection in intensive care units. *JAMA*. 2009;302(21):2323–9.
- Kuti EL, Patel AA, Coleman CI. Impact of inappropriate antibiotic therapy on mortality in patients with ventilator-associated pneumonia and blood stream infection: a meta-analysis. *J Crit Care*. 2008;23(1):91–100.
- Melsen WG, Rovers MM, Groenwold RH, Bergmans DC, Camus C, Bauer TT, et al. Attributable mortality of ventilator-associated pneumonia: a meta-analysis of individual patient data from randomised prevention studies. *Lancet Infect Dis*. 2013;13(8):665–71.
- Klompas M. Eight initiatives that misleadingly lower ventilator-associated pneumonia rates. *Am J Infect Control*. 2012;40(5):408–10.
- Grgurich PE, Hudcova J, Lei Y, Sarwar A, Craven DE. Diagnosis of ventilator-associated pneumonia: controversies and working toward a gold standard. *Curr Opin Infect Dis*. 2013;26(2):140–50.
- Novosel TJ, Hodge LA, Weireter LJ, Britt RC, Collins JN, Reed SF, et al. Ventilator-associated pneumonia: depends on your definition. *Am Surg*. 2012;78(8):851–4.
- Raof S, Baumann MH. An official multi-society statement: ventilator-associated events: the new definition. *Crit Care Med*. 2014;42(1):228–9.
- Grossman RF, Fein A. Evidence-based assessment of diagnostic tests for ventilator-associated pneumonia. Executive Summary *Chest*. 2000;117(4 Suppl 2):1775–815.
- Hunter JD. Ventilator associated pneumonia. *BMJ*. 2012;344, e3325.
- Rello J, Diaz E, Rodriguez A. Etiology of ventilator-associated pneumonia. *Clin Chest Med*. 2005;26(1):87–95.
- Cabello H, Torres A, Celis R, El-Ebiary M, Puig de la Bellacasa J, Xaubet A, et al. Bacterial colonization of distal airways in healthy subjects and chronic lung disease: a bronchoscopic study. *Eur Respir J*. 1997;10(5):1137–44.
- American Thoracic S, Infectious Diseases Society of A. Guidelines for the management of adults with hospital-acquired, ventilator-associated, and healthcare-associated pneumonia. *Am J Respir Crit Care Med*. 2005;171(4):388–416.
- Carratala J, Roson B, Fernandez-Sevilla A, Alcaide F, Gudiol F. Bacteremic pneumonia in neutropenic patients with cancer: causes, empirical antibiotic therapy, and outcome. *Arch Intern Med*. 1998;158(8):868–72.
- Shenep JL. Viridans-group streptococcal infections in immunocompromised hosts. *Int J Antimicrob Agents*. 2000;14(2):129–35.
- Wong CA, Donald F, Macfarlane JT. Streptococcus milleri pulmonary disease: a review and clinical description of 25 patients. *Thorax*. 1995;50(10):1093–6.
- Shinzato T, Saito A. The Streptococcus milleri group as a cause of pulmonary infections. *Clin Infect Dis*. 1995;21 Suppl 3:S238–43.
- Hauser AR, Jain M, Bar-Meir M, McColley SA. Clinical significance of microbial infection and adaptation in cystic fibrosis. *Clin Microbiol Rev*. 2011;24(1):29–70.
- Sibley CD, Parkins MD, Rabin HR, Duan K, Norgaard JC, Surette MG. A polymicrobial perspective of pulmonary infections exposes an enigmatic pathogen in cystic fibrosis patients. *Proc Natl Acad Sci U S A*. 2008;105(39):15070–5.
- DiBardino DM, Wunderink RG. Aspiration pneumonia: A review of modern trends. *J Crit Care*. 2015;30(1):40–8.
- Opal SM. Immunologic alterations and the pathogenesis of organ failure in the ICU. *Semin Respir Crit Care Med*. 2011;32(5):569–80.
- Kox M, Vrouwenvelder MQ, Pompe JC, van der Hoeven JG, Pickkers P, Hoedemaekers CW. The effects of brain injury on heart rate variability and the innate immune response in critically ill patients. *J Neurotrauma*. 2012;29(5):747–55.
- Conway Morris A, Anderson N, Brittan M, Wilkinson TS, McAuley DF, Antonelli J, et al. Combined dysfunctions of immune cells predict nosocomial infection in critically ill patients. *Br J Anaesth*. 2013;111(5):778–87.
- Bonten MJ, Bergmans DC, Stobberingh EE, van der Geest S, De Leeuw PW, van Tiel FH, et al. Implementation of bronchoscopic techniques in the diagnosis of ventilator-associated pneumonia to reduce antibiotic use. *Am J Respir Crit Care Med*. 1997;156(6):1820–4.
- Linssen CF, Jacobs JA, Schouten JS, van Mook WN, Ramsay G, Drent M. Influence of antibiotic therapy on the cytological diagnosis of ventilator-associated pneumonia. *Intensive Care Med*. 2008;34(5):865–72.
- File TM. Community-acquired pneumonia. *Lancet*. 2003;362(9400):1991–2001.
- Murray PR, Rosenthal KS, Pfäller MA, editors. *Medical Microbiology*. Seventh Edition. Elsevier; 2012. ISBN: 978-0-323-08692-9 [http://www.elsevier.com/books/medical-microbiology/murray/978-0-323-08692-9].
- el-Ebiary M, Torres A, Fabregas N, de la Bellacasa JP, Gonzalez J, Ramirez J, et al. Significance of the isolation of *Candida* species from respiratory samples in critically ill, non-neutropenic patients. An immediate postmortem histologic study. *Am J Respir Crit Care Med*. 1997;156(2 Pt 1):583.
- Magill SS, Klompas M, Balk R, Burns SM, Deutschman CS, Diekema D, et al. Developing a new, national approach to surveillance for ventilator-associated events*. *Crit Care Med*. 2013;41(11):2467–75.

31. Medford AR, Husain SA, Turki HM, Millar AB. Diagnosis of ventilator-associated pneumonia. *J Crit Care*. 2009;24(3):473. e471-476.
32. Canadian Critical Care Trials G. A randomized trial of diagnostic techniques for ventilator-associated pneumonia. *N Engl J Med*. 2006;355(25):2619-30.
33. Knaus WA, Draper EA, Wagner DP, Zimmerman JE. APACHE II: a severity of disease classification system. *Crit Care Med*. 1985;13(10):818-29.
34. Vincent JL, de Mendonca A, Cantraine F, Moreno R, Takala J, Suter PM, et al. Use of the SOFA score to assess the incidence of organ dysfunction/failure in intensive care units: results of a multicenter, prospective study. Working group on "sepsis-related problems" of the European Society of Intensive Care Medicine. *Crit Care Med*. 1998;26(11):1793-800.
35. Ferreira FL, Bota DP, Bross A, Melot C, Vincent JL. Serial evaluation of the SOFA score to predict outcome in critically ill patients. *JAMA*. 2001;286(14):1754-8.
36. Pugin J, Auckenthaler R, Mili N, Janssens JP, Lew PD, Suter PM. Diagnosis of ventilator-associated pneumonia by bacteriologic analysis of bronchoscopic and nonbronchoscopic "blind" bronchoalveolar lavage fluid. *Am Rev Respir Dis*. 1991;143(5 Pt 1):1121-9.
37. De Brauwere EI, Jacobs JA, Nieman F, Bruggeman CA, Wagenaar SS, Drent M. Cytocentrifugation conditions affecting the differential cell count in bronchoalveolar lavage fluid. *Anal Quant Cytol Histol*. 2000;22(5):416-22.
38. Clark AE, Kaleta EJ, Arora A, Wolk DM. Matrix-assisted laser desorption ionization-time of flight mass spectrometry: a fundamental shift in the routine practice of clinical microbiology. *Clin Microbiol Rev*. 2013;26(3):547-603.
39. Lambotte O, Timsit JF, Garrouste-Orgeas M, Misset B, Benali A, Carlet J. The significance of distal bronchial samples with commensals in ventilator-associated pneumonia: colonizer or pathogen? *Chest*. 2002;122(4):1389-99.
40. Rouby JJ, Martin De Lassale E, Poete P, Nicolas MH, Bodin L, Jarlier V, et al. Nosocomial bronchopneumonia in the critically ill. Histologic and bacteriologic aspects. *Am Rev Respir Dis*. 1992;146(4):1059-66.
41. Kirtland SH, Corley DE, Winterbauer RH, Springmeyer SC, Casey KR, Hampson NB, et al. The diagnosis of ventilator-associated pneumonia: a comparison of histologic, microbiologic, and clinical criteria. *Chest*. 1997;112(2):445-57.
42. Torres A, el-Ebiary M, Padro L, Gonzalez J, de la Bellacasa JP, Ramirez J, et al. Validation of different techniques for the diagnosis of ventilator-associated pneumonia. Comparison with immediate postmortem pulmonary biopsy. *Am J Respir Crit Care Med*. 1994;149(2 Pt 1):324-31.
43. Papazian L, Thomas P, Garbe L, Guignon I, Thirion X, Charrel J, et al. Bronchoscopic or blind sampling techniques for the diagnosis of ventilator-associated pneumonia. *Am J Respir Crit Care Med*. 1995;152(6 Pt 1):1982-91.
44. Van de Voort PHJ, van Saene HKF, editors. *Selective Digestive Tract Decontamination in Intensive Care Medicine. A Practical Guide to Controlling Infection*. 1st Edition. Italy: Springer-Verlag Italia; 2008.
45. Schnabel RM, Scholte JB, Van Der Velden KE, Roekaerts PM, Bergmans DC. Ventilator-associated pneumonia rates after introducing selective digestive tract decontamination. *Infect Dis*. 2015;1-4.
46. Scholte JB, van Dessel HA, Linssen CF, Bergmans DC, Savelkoul PH, Roekaerts PM, et al. Endotracheal aspirate and bronchoalveolar lavage fluid analysis: interchangeable diagnostic modalities in suspected ventilator-associated pneumonia? *J Clin Microbiol*. 2014;52(10):3597-604.
47. Kramer A, Schwabke I, Kampf G. How long do nosocomial pathogens persist on inanimate surfaces? A systematic review *BMC Infect Dis*. 2006;6:130.
48. Pileggi C, Bianco A, Flotta D, Nobile CG, Pavia M. Prevention of ventilator-associated pneumonia, mortality and all intensive care unit acquired infections by topically applied antimicrobial or antiseptic agents: a meta-analysis of randomized controlled trials in intensive care units. *Crit Care*. 2011;15(3):R155.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

