

RESEARCH

Open Access



Characteristics and prognostic implications of peripheral blood lymphocyte subsets in patients with anti-MDA5 antibody positive dermatomyositis-interstitial lung disease

Fang-Ping Ren¹, Qi Chen¹, Shan-Shan Yao², Lin Feng³, Xin-Ying Xue¹, Wei-Chao Zhao⁴, Dong Wang³, Zhi-Ling Zhao³, Si-Wei Gu⁵, Ting Li³, Ya-Wen Shen¹, Lan Gao¹, Xue-Lei Zang¹, Xin-Yu Bao⁶ and Zhao-Hui Tong^{3*}

Abstract

Objectives To examine the characteristics of blood lymphocyte subsets in dermatomyositis-interstitial lung disease (DM-ILD) inflicted patients with positive anti-melanoma differentiation-associated gene 5 (anti-MDA5), as well as its prognosis value in this set of patients.

Methods Data were retrospectively collected from 253 DM-ILD patients from three hospitals in China between January 2016 to January 2021. Patients were grouped into anti-MDA5 antibody positive group (MDA5⁺ DM-ILD) and anti-MDA5 antibody negative group (MDA5⁻ DM-ILD) based on myositis-specific autoantibody test results. Demographic characteristics, lymphocyte subsets patterns and other clinical features were compared between the two groups. The association of lymphocyte subsets with 180-day mortality was investigated using survival analysis in MDA5⁺ DM-ILD.

Results Out of 253 eligible patients with DM-ILD, 59 patients were anti-MDA5⁺ and 194 were anti-MDA5⁻. Peripheral blood lymphocyte count, CD3⁺ count, percentage of CD3⁺, CD3⁺CD4⁺ count, and CD3⁺CD8⁺ count was lower in MDA5⁺ DM-ILD than in MDA5⁻ DM-ILD (all $P < 0.001$) as well as CD3⁺CD19⁺ count ($P = 0.04$). In MDA5⁺ DM-ILD, CD3⁺CD8⁺ count ≤ 49.22 cell/ μ L (HR = 3.81, 95%CI [1.20, 12.14]) and CD3⁺CD19⁺ count ≤ 137.64 cell/ μ L (HR = 3.43, 95%CI [1.15, 10.24]) were independent predictors of mortality. CD3⁺CD8⁺ count ≤ 31.38 cell/ μ L was associated with a higher mortality risk in all DM-ILD patients (HR = 8.6, 95%CI [2.12, 31.44]) after adjusting for anti-MDA5 and other clinical characteristics.

Conclusion Significant lymphocytes decrease was observed in MDA5⁺ DM-ILD patients. CD3⁺CD8⁺ cell count was associated with worse prognosis in both MDA5⁺ DM-ILD and all DM-ILD patients.

Keywords Anti-melanoma differentiation-associated gene 5, Dermatomyositis, Interstitial lung disease, Lymphocyte subsets, Prognostic

*Correspondence:

Zhao-Hui Tong
tongzhaohuicy@sina.com

Full list of author information is available at the end of the article



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Introduction

Dermatomyositis (DM) is an idiopathic inflammatory disease that causes muscle weakness and skin rashes. DM patients present different phenotypes and clinical courses that could be complicated with interstitial lung disease (ILD), which is associated with poor prognosis [1, 2]. A variety of myositis-specific antibodies (MSAs) had been identified for phenotyping of DM and early recognition of high risk patients, such as the most prevalent anti-Jo-1 (occurring in 9–24% of adult DM patients [3]), anti-melanoma differentiation-associated gene 5 (anti-MDA5, occurring from 15 to 20% in Asian DM patients [4]), and less common anti-PL-7, anti-EJ, anti-PL-12, anti-OJ, et al. [5, 6]. Anti-MDA5 has drawn increasing attention due to the high occurrence rate of ILD in anti-MDA5⁺ DM (MDA5⁺DM), among 50–70% [7, 8]. Anti-MDA5⁺ DM-ILD (MDA5⁺ DM-ILD) is associated with rapid progressive ILD, glucocorticoid resistance and often fatal outcomes [9–11].

The autoimmune mechanisms underlying MDA5⁺ DM-ILD are poorly understood [12–14]. Previous researches were mainly conducted in MDA5⁺ DM and showed that lymphocyte infiltration was involved in this pathogenesis [15]. Lymphocytes recruitment was found in the lung in MDA5⁺ DM patients and the circulatory lymphocytes were diminished, including the subsets T lymphocytes and B lymphocytes [16–18]. Lymphocytes targeted therapeutic has proved effective in the treatment of MDA5⁺ DM [19–21]. Further research of the immunological cellular characteristics in MDA5⁺ DM-ILD might help to understand the autoimmune mechanism underlying this high-risk subgroup and shed light to therapeutic methods.

Here we examined the immunological cellular characteristics in MDA5⁺ DM-ILD and explored possible prognostic factors.

Method

Patients

A total of 253 patients with DM who were diagnosed with ILD in the Department of Respiratory Medicine and the Department of Rheumatology and Immunology at Beijing Chao-Yang Hospital, Capital Medical University, Beijing Shijitan Hospital, Capital Medical University, and PLA Strategic Support Force Medical Center from January 1, 2016 to January 1, 2021 were included in this study. Demographic and medical records were obtained from the Electronic Medical Records (EMR) system. We recorded age, sex, smoking history, chronic disease, blood test results, lymphocyte subsets, MSAs spectrum, and survival status upon discharge. We conducted telephone follow-up 180 days after discharge.

Inclusion and exclusion criteria

Inclusion criteria:

1. Aged between 18 and 80;
2. Compliance with the DM diagnostic criteria recommended by Bohan/Peter [22, 23] or Sontheimer's proposed CADM criteria (1999) [24];
3. With ILD manifestations identified by chest HRCT;
4. With positive MSAs demonstrated by myositis antibody spectrum assay prior to treatment;
5. With complete test results of peripheral blood lymphocyte subsets present prior to treatment.

Exclusion criteria:

1. History of tumor or chronic lung disease;
2. Complicated by other connective tissue diseases, such as systemic sclerosis (SSc), Sjögren's syndrome (SS), rheumatoid arthritis (RA), and systemic lupus erythematosus (SLE);
3. Received systemic glucocorticoid and immunosuppressant treatment prior to hospitalization.

All patients were anonymized. Based on EMR, 902 patients with DM-ILD were included from 1,573 patients with idiopathic inflammatory myopathy (IIM). After excluding 163 patients based on exclusion criteria, 253 patients had laboratory results for lymphocyte subsets and positive MSAs, including 59 patients with anti-MDA5 positive (MDA5⁺) and 194 patients with anti-MDA5 negative (MDA5⁻), were included in the analysis (Fig. 1).

DM Serotyping (MSAs)

MSAs were assayed using WESTERN blotting. We defined MDA5⁺ DM-ILD as DM-ILD with positive anti-MDA5 antibody. Positivity for other MSAs was recorded.

Imaging analysis

High-resolution computed tomography (HRCT) images were obtained from Picture Archiving and Communication System (PACS) in study centers. Patients with ground-glass opacity, cord-like and reticular fiber, and/or consolidation on chest HRCT were diagnosed as ILD. According to imaging and pathological characteristics, ILD was classified into usual interstitial pneumonia (UIP), nonspecific interstitial pneumonia (NSIP), organizing pneumonia (OP), diffuse alveolar damage (DAD), and mixed NSIP-OP. Mixed NSIP-OP is distinguished by a predominately basal fibrotic abnormality with superimposed OP [25]. The ILD diagnosis was confirmed through HRCT and weekly discussion by a multidisciplinary team (MDT) consisting of two pulmonary physicians specializing in interstitial lung disease, one rheumatologist, two radiologists, one pathologist, and one internist. If the MDT had a high level of confidence in radiological diagnosis ($\geq 90\%$) and if HRCT radiological characteristics

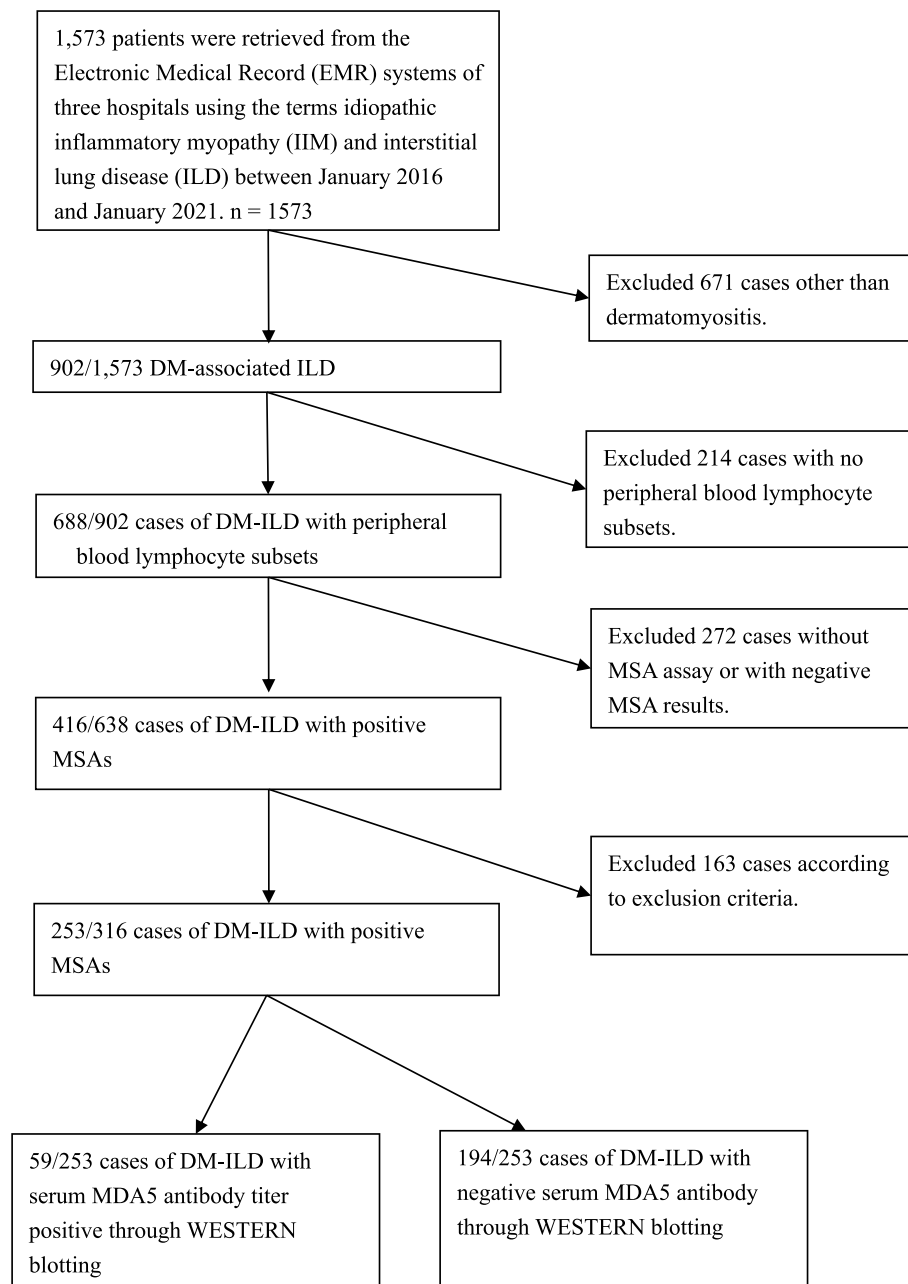


Fig. 1 Sample selection profile. This flowchart shows how 253 DM-ILD patients were selected. The flowchart has six steps: Search EMR system for IIM with ILD. Exclude non-DM-ILD. Exclude the patients with no blood lymphocyte or MSA profile. Exclude the patients that met other exclusion criteria. Divide the remaining participants into MDA5⁺ DM-ILD and MDA5⁻ DM-ILD and enroll in the final cohort

were typical, the MDT agreed that the type of ILD could be confirmed without a biopsy. In the case of atypical HRCT findings or objections to the pathological classification of ILD, the MDT engaged in deliberation. Rapidly progressive interstitial lung disease (RPILD) was defined as worsening radiological interstitial change with progressive dyspnea and hypoxemia within one month of the onset of respiratory symptoms [26].

Lymphocyte subsets

Peripheral blood lymphocyte subsets were tested at the first visit with flow cytometry assay including counts and proportions of T lymphocytes (CD3⁺), helper/inducer T lymphocytes (CD3⁺CD4⁺ cells), suppressive/cytotoxic T lymphocytes (CD3⁺CD8⁺ cells), B lymphocytes (CD3⁻CD19⁺ cells), and NK cells (CD3⁻CD56⁺ cells).

Statistical analysis

Categorical variables were summarized as frequency (proportion). Chi-squared test or the Fisher's exact test were utilized to compare proportions between groups, as appropriate. Continuous variables were presented as mean (standard deviation) or median (25th percentile, 75th percentile). Normality of continuous variables were examined using Kolmogorov–Smirnov test. The Kolmogorov–Smirnov test was also used to compare continuous variables between groups as all variables rejected normality. The cutoff thresholds for continuous data were determined using the "survminer" R package and the maximum selection log-rank test. The Cox proportional hazard model was used to estimate the hazard ratios (HRs)

Table 1 Baseline characteristics in MDA5⁺ DM-ILD and MDA5⁻ DM-ILD patients

Baseline	MDA5 ⁺ DM-ILD cohort (n = 59)	MDA5 ⁻ DM-ILD cohort (n = 194)	P value
Age (years)	51 (46.5, 62.5)	57 (48.0, 64.0)	0.106
Female	37 (62.7%)	136 (70.1%)	0.285
Smoking history	11 (18.6%)	31 (16.0%)	0.63
180-day all-cause death	30 (50.8%)	15 (7.7%)	<0.001

Table 1 compares the baseline characteristics of patients with DM-ILD who were positive or negative for MDA5 antibodies. The study included 59 MDA5⁺ DM-ILD patients and 194 MDA5⁻ DM-ILD patients. There were no significant differences between the two groups in terms of age, sex, and smoking history. However, the MDA5⁺ DM-ILD group had a significantly higher 180-day all-cause mortality rate than the MDA5⁻ DM-ILD group (50.8% vs. 7.7%, $P < 0.001$)

$P < 0.05$ is in bold

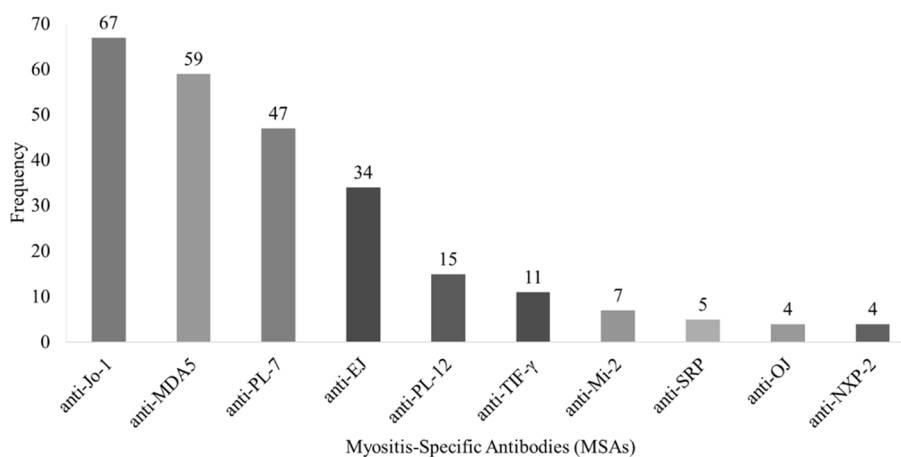


Fig. 2 Distribution of MSAs among all DM-ILD patients ($N = 253$). Abbreviations: anti-Jo-1, anti-histidyl-tRNA synthetase; anti-MDA5, anti-melanoma differentiation-associated gene 5; anti-PL-7, anti-threonyl-tRNA synthetase; anti-EJ, anti-glycyl-tRNA synthetase; anti-PL-12, anti-alanyl-tRNA synthetase; anti-TIF1 γ , anti-transcription intermediary factor-1 γ ; anti-Mi-2, anti-complex nucleosome remodeling histone deacetylase; anti-SRP, anti-signal recognition particle; anti-OJ, anti-isoleucyl-tRNA synthetase; anti-NXP-2, anti-nuclear matrix protein-2. Figure shows the distribution of MSAs in all DM-ILD patients. The most prevalent antibody was anti-Jo-1 (67 cases, 26.5%), followed by: anti-MDA5 (59 cases, 23.3%), anti-PL-7 (47 cases, 18.6%) and anti-EJ (34 cases, 13.4%). Other detected antibodies included anti-PL-12, anti-TIF1 γ , anti-Mi-2, anti-SRP, anti-OJ and anti-NXP-2

and corresponding 95% Confidence Interval (95%CI) for all-cause mortality within 180 days, with the proportional hazard hypothesis investigated using Schoenfeld residuals. All analyses were performed using R Project for Statistical Computing (version 4.2.1). The two-sided P values < 0.05 were considered statistically significant.

Results

Baseline characteristics in MDA5⁺ DM-ILD and MDA5⁻ DM-ILD patients

From January 2016 to January 2021, 253 patients of DM-ILD were screened in the study, of whom 59 were MDA5⁺ DM-ILD and 194 were MDA5⁻ DM-ILD according to the inclusion criteria (Table 1). The median age of all patients was 55 years, with 68.4% being females. There was no significant difference in age and sex distribution between MDA5⁺ DM-ILD and MDA5⁻ DM-ILD patients ($P = 0.106$). During the 180-day follow-up, 30 (50.8%) deaths among MDA5⁺ patients and 15 (7.7%) were recorded among MDA5⁻ patients, which was significantly different ($P < 0.001$).

Figure 2 depicts the distribution of MSAs from both cohorts. Among 253 patients with DM-ILD, 59 (23.3%) patients were anti-MDA5 positive. The majority of the 194 anti-MDA5⁻ patients were positive for anti-Jo-1 ($n = 67$), anti-PL-7 ($n = 47$), and anti-EJ ($n = 34$).

MDA5⁺ DM-ILD exhibits more aggressive clinical traits

Lung imaging diagnosis results and relevant clinical characteristics are presented in Table 2. DAD was the most prevalent in MDA5⁺ DM-ILD patients, whereas

Table 2 Imaging study and clinical traits of patients in MDA5⁺ DM-ILD and MDA5⁻ DM-ILD cohorts

Characteristic	MDA5 ⁺ DM-ILD (n = 59)	MDA5 ⁻ DM-ILD (n = 194)	P value
ILD pattern (Radiological and pathological diagnosis by MDT)			
NSIP	20(33.9)	150(77.3)	<0.001
OP	12(20.3)	21(10.8)	0.057
UIP	3(5.1)	5(2.6)	0.394
DAD	20(33.9)	1(0.5)	<0.001
mixed NSIP-OP	4(6.8)	17(8.8)	0.791
RPILD	29(49.2)	10(5.2)	<0.001
length of hospital stay (d)	14(8.00, 19.50)	12(8.00, 14.75)	0.005

Table 2 shows the differences in clinical, imaging, and pathological features between MDA5⁺ DM-ILD and MDA5⁻ DM-ILD. MDA5⁺ DM-ILD patients had a significantly lower rate of NSIP, and a significantly higher rate of DAD. MDA5⁺ DM-ILD patients also had longer hospital stays than MDA5⁻ DM-ILD patients

P < 0.05 are in bold

Abbreviations: NSIP nonspecific interstitial pneumonia, OP organizing pneumonia, UIP usual interstitial pneumonia, DAD diffuse alveolar damage, RPILD rapidly progressive interstitial lung disease

NSIP was the most prevalent in MDA5⁻ DM-ILD patients. RPILD was observed in 49.2% of patients with MDA5⁺ DM-ILD (*n* = 29) compared to 5.2% in patients with MDA5⁻ DM-ILD (*n* = 10) (*P* < 0.001). Patients with MDA5⁺ DM-ILD had a longer average length of hospital stay (*P* = 0.005).

MDA5⁺ DM-ILD Features more intensive lymphocyte depletion and activation

Comparison of peripheral blood lymphocyte subsets between MDA5⁺ DM-ILD and MDA5⁻ DM-ILD patients are presented in Table 3. The total number of lymphocytes in peripheral blood was significantly lower (*P* < 0.001) in MDA5⁺ patients than in MDA5⁻. In the MDA5⁺ DM-ILD patients, the percentage and count of CD3⁺ cells were significantly lower (both *P* < 0.001), as

did the count of CD3⁻ CD19⁺ cells (*P* = 0.04). Analysis of subtypes of T-lymphocytes revealed a lower count of all subtypes (all *P* < 0.001). Sub-analysis of lymphocyte subsets in major types of MSAs DM-ILD (> 10% of total study population) showed a significantly lower total lymphocyte count, CD3⁺ cell count and CD3⁺CD4⁺ cell count (all *P* < 0.05) (Table 4).

CD3⁺CD8⁺ Count and CD3⁻CD19⁺ count predict mortality in MDA5⁺ DM-ILD

We compared lymphocyte subsets between survived and deceased patients in the MDA5⁺ DM-ILD cohort (Supplementary material 1). CD3⁺CD8⁺ count and CD3⁺CD19⁺ count was identified as significant predictors of mortality (Fig. 3A, B). Kaplan–Meier survival curves demonstrated statistically significant differences

Table 3 Peripheral blood lymphocyte subsets of patients in MDA5⁺ DM-ILD and MDA5⁻ DM-ILD cohorts

Lymphocyte subset	MDA5 ⁺ DM-ILD (n = 59)	MDA5 ⁻ DM-ILD (n = 194)	P value
blood lymphocyte count (× 10 ⁹ /L)	0.66(0.46, 0.96)	1.075(0.78, 1.43)	<0.001
CD3 ⁺ (%)	60.3(54.85, 64.85)	65.695(54.92, 75.58)	<0.001
CD3 ⁺ (cell/μL)	402.97(248.18, 553.37)	659.5(445.75, 919.00)	<0.001
CD3 ⁺ CD4 ⁺ (%)	38.6(35.51, 44.85)	42.1(34.10, 48.58)	0.077
CD3 ⁺ CD4 ⁺ (cell/μL)	255.51(171.55, 369.18)	433.5(275.25, 592.50)	<0.001
CD3 ⁺ CD8 ⁺ (%)	19.1(15.28, 22.99)	20.23(14.96, 26.78)	0.085
CD3 ⁺ CD8 ⁺ (cell/μL)	123(72.71, 200.48)	205.5(129.00, 316.50)	<0.001
CD4 ⁺ /CD8 ⁺	1.98(1.67, 2.75)	2.025(1.39, 2.85)	0.346
CD3 ⁻ CD19 ⁺ (%)	18.5(14.15, 25.28)	17.38(9.85, 29.21)	0.037
CD3 ⁻ CD19 ⁺ (cell/μL)	119.38(77.06, 191.95)	165.44(89.38, 313.84)	0.04
CD3 ⁻ CD56 ⁺ (%)	12.7(9.80, 16.45)	9.3(6.00, 13.93)	<0.001
CD3 ⁻ CD56 ⁺ (cell/μL)	86.13(55.58, 134.70)	96.87(54.99, 155.80)	0.701

Table 3 compares the lymphocyte subsets of patients with DM-ILD according to their anti-MDA5 antibody status. The MDA5⁺ DM-ILD group had significantly lower counts of total lymphocytes, CD3⁺ cells, CD3⁺CD4⁺ cells, CD3⁺CD8⁺ cells, and CD3⁻CD19⁺ cells than the MDA5⁻ DM-ILD group. The MDA5⁺ DM-ILD group also had a lower percentage of CD3⁺ cells, but higher percentages of CD3⁻CD56⁺ cells and CD3⁻CD19⁺ cells than the MDA5⁻ DM-ILD group

P < 0.05 are in bold

Table 4 Peripheral blood lymphocyte subsets of patients with MDA5⁺ DM-ILD and other MSAs positive DM-ILD

Lymphocyte subset	MDA5 ⁺ DM-ILD (n = 59)	anti-Jo-1 ⁺ DM-ILD (n = 71)	anti-PL-7 ⁺ DM-ILD (n = 47)	anti-EJ ⁺ DM-ILD (n = 34)
blood lymphocyte count (× 10 ⁹ /L)	0.66(0.46, 0.96)	1.12(0.83, 1.53) ***	1.08(0.81, 1.44) ***	1(0.74, 1.32) **
CD3 ⁺ (%)	60.3(54.85, 64.85)	69.3(58.22, 77.98) ***	66.2(56.72, 74.75) **	58.9(51.85, 70.85)
CD3 ⁺ (cell/μL)	402.97(248.18, 553.37)	663(450.5, 980.5) ***	695(529.5, 914.74) ***	586(396, 917.1) *
CD3 ⁺ CD4 ⁺ (%)	38.6(35.51, 44.85)	41.02(35.92, 50.18)	43.01(36.45, 48) *	42.86(34.15, 47.5)
CD3 ⁺ CD4 ⁺ (cell/μL)	255.51(171.55, 369.18)	448(280.5, 651.5) ***	458(315,586) ***	387(243, 587.5) *
CD3 ⁺ CD8 ⁺ (%)	19.1(15.28, 22.99)	21.7(16.4, 27.2) *	21.5(14.95, 27.99)	17.49(12.3, 23.44)
CD3 ⁺ CD8 ⁺ (cell/μL)	123(72.71, 200.48)	227(133, 334.5) ***	231(158.5, 301) ***	168(119, 272.5)
CD4 ⁺ /CD8 ⁺	1.98(1.67, 2.75)	1.94(1.48, 2.71)	1.98(1.29, 2.95)	2.26(1.35, 3.28)
CD3 ⁻ CD19 ⁺ (%)	18.5(14.15, 25.28)	13.61(9.7, 26.95) **	16.46(8.95, 26.86)	18.3(8.5, 30.7)
CD3 ⁻ CD19 ⁺ (cell/μL)	119.38(77.06, 191.95)	152.4(75.77, 317.2)	176.3(86.91, 289.85)	166.95(58.31, 269.69)
CD3 ⁻ CD56 ⁺ (%)	12.7(9.80, 16.45)	9(5.6, 12.9) ***	9(6.01, 13) ***	12.8(6.43, 16.6)
CD3 ⁻ CD56 ⁺ (cell/μL)	86.13(55.58, 134.70)	79.52(44.8, 151.12)	105(61.2, 150.2)	113.12(72.74, 182.16)

Table 4 compares the lymphocyte subsets between MDA5⁺ DM-ILD, anti-Jo-1⁺ DM-ILD, anti-PL-7⁺ DM-ILD and anti-EJ⁺ DM-ILD. The MDA5⁺ DM-ILD group had significantly lower counts of total lymphocytes, CD3⁺ cells, CD3⁺CD4⁺ T cells compared to other groups of MSAs DM-ILD

* P value < 0.05 compared with MDA5⁺ DM-ILD. ** P value < 0.01 compared with MDA5⁺ DM-ILD. *** P value < 0.001 compared with MDA5⁺ DM-ILD

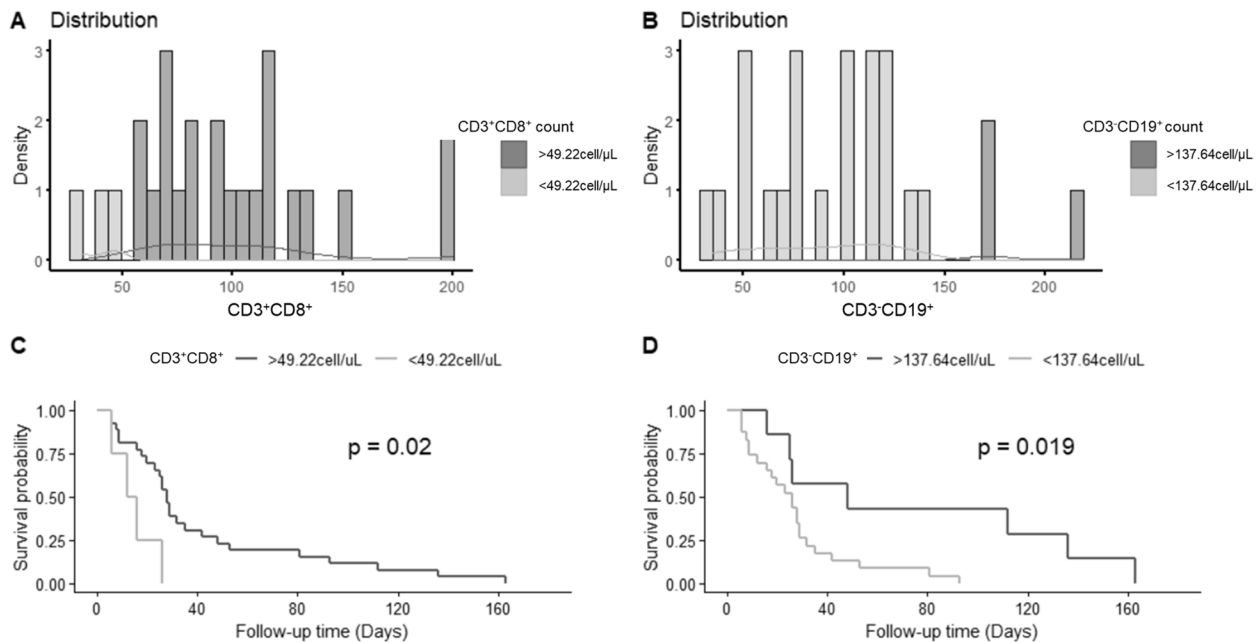


Fig. 3 **A, B** Optimal cut-off values for CD3⁺CD8⁺ count and CD3⁻CD19⁺ count using the “survminer” R package. **C, D** The Kaplan–Meier survival curves displaying 180-day all-cause mortality, based on CD3⁺CD8⁺ count (with cut-off value of 49.22 cell/μL) and CD3⁻CD19⁺ count (with cut-off value of 137.64 cell/μL). Figure shows how the survival outcome of patients with MDA5⁺ DM-ILD is related to the counts of two types of lymphocytes: CD3⁺CD8⁺ T cells and CD3⁻CD19⁺ B cells. The cut-off value for CD3⁺CD8⁺ T cells is 49.22 cell/μL and the cut-off value for CD3⁻CD19⁺ B cells is 137.64 cell/μL. The histograms in panels A and C show the number of patients with recorded outcomes in each group. The Kaplan–Meier curves in panels B and D show the survival probability of each group over time. The log-rank tests show that both lymphocyte counts are significantly associated with survival outcome. Patients with higher CD3⁺CD8⁺ T cell count or higher CD3⁻CD19⁺ B cell count have a better prognosis than those with lower CD3⁺CD8⁺ T cell count or lower CD3⁻CD19⁺ B cell count. The P-values for the log-rank tests are 0.02 and 0.019, respectively

between groups (Fig. 3C, D, all $P < 0.05$). Table 5 displays the Cox regression hazard ratios (HR).

Anti-MDA5 positivity and CD3⁺CD8⁺ count independently predict mortality in all patients with DM-ILD

Anti-MDA5 positivity was associated with higher mortality (HR = 2.08[1.64,13.22], $P = 0.032$) among all patients. CD3⁺CD8⁺ ≤ 31.38 cell/ μ L was associated with 180-day mortality (HR = 8.6[2.12,31.44], $P = 0.002$) after adjusting for sex, age, MDA5 status and RPILD (Table 6, Fig. 4A, B).

Discussion

In this study, we examined 253 patients with DM-ILD, the largest number of participants to date. Specifically, Jo-1 was the most prevalent antibody, followed by MDA5 and PL-7, with MDA5 having a 23.3% positive rate (Fig. 2). The distribution of myositis antibodies in Asian populations was essentially consistent with previous reports [27]. Our study covered 59 patients with MDA5⁺ DM-ILD. This is the largest study to date in terms of the number of MDA5⁺ DM-ILD participants.

Compared to previous studies that focused on DM-ILD as a whole and with non-ILD DM patients as controls

Table 5 Hazard ratio (HR) for prognostic factors in MDA5⁺ DM-ILD

Factor	Adjusted HR(95%CI)	P value
CD3 ⁺ cell count (< 233.12 cell/ μ L)	1.36(0.59,3.12)	0.47
CD3 ⁺ % (< 53.29%)	0.61(0.22,1.71)	0.35
CD3 ⁺ CD4 ⁺ cell count (< 193.5 cell/ μ L)	1.41(0.63,3.13)	0.4
CD3 ⁺ CD4 ⁺ % (< 38.7%)	0.44(0.18,1.04)	0.06
CD3 ⁺ CD8 ⁺ cell count (< 49.22 cell/ μ L)	3.81(1.20,12.14)	0.023
CD3 ⁺ CD8 ⁺ % (< 10.7%)	4.56(0.87,23.94)	0.073
CD3 ⁺ CD4 ⁺ cell count/CD3 ⁺ CD8 ⁺ cell count (< 2.99)	0.37(0.13,1.06)	0.06
CD3 ⁻ CD19 ⁺ cell count (< 137.64 cell/ μ L)	3.43(1.15,10.24)	0.027
CD3 ⁻ CD19 ⁺ % (< 27%)	3.45(0.84,14.19)	0.087
CD3 ⁻ CD56 ⁺ cell count (< 67.62 cell/ μ L)	2.84(0.98,23.46)	0.052
CD3 ⁻ CD56 ⁺ % (< 8.2%)	4.89(0.97,24.72)	0.055

Table 5 shows the results of the multivariate Cox regression analysis for MDA5⁺ DM-ILD survival. The analysis identified two independent prognostic factors that were significantly associated with increased mortality risk: CD3⁺CD8⁺ cell count lower than 49.22 cell/ μ L (HR = 3.81, 95% CI = 1.20-12.14, $P = 0.023$) and CD3⁻CD19⁺ cell count lower than 137.64 cell/ μ L (HR = 3.43, 95% CI = 1.15-10.24, $P = 0.027$)

$P < 0.05$ are in bold

Table 6 Hazard ratio (HR) for prognostic factors in all DM-ILD

Factor	Adjusted HR(95%CI)	P value
Anti-MDA5 positive	2.08(1.64,13.22)	0.032
CD3 ⁺ cell count (< 420.29 cell/ μ L)	2.18(1.4,63)	0.052
CD3 ⁺ % (< 56.07%)	0.81(0.42,1.56)	0.53
CD3 ⁺ CD4 ⁺ cell count (< 193.5 cell/ μ L)	1.68(0.9,3.15)	0.61
CD3 ⁺ CD4 ⁺ % (< 36.15%)	0.85(0.41,1.76)	0.7
CD3 ⁺ CD8 ⁺ cell count (< 31.38 cell/ μ L)	8.6(2.12,31.44)	0.002
CD3 ⁺ CD8 ⁺ % (< 10.7%)	6.78(1.74,26.53)	0.006
CD3 ⁺ CD4 ⁺ cell count/CD3 ⁺ CD8 ⁺ cell count (< 2.97)	0.7(0.34,1.42)	0.32
CD3 ⁻ CD19 ⁺ cell count (< 137.64 cell/ μ L)	2.02(0.97,4.24)	0.062
CD3 ⁻ CD19 ⁺ % (< 27%)	2.3(0.91,5.1)	0.12
CD3 ⁻ CD56 ⁺ cell count (< 40.32 cell/ μ L)	1.74(0.88,3.44)	0.11
CD3 ⁻ CD56 ⁺ % (< 7.5%)	2.17(0.75,6.28)	0.15

Table 6 shows the results of the multivariate Cox regression analysis for DM-ILD survival. The analysis identified two independent prognostic factors that were significantly associated with increased mortality risk: anti-MDA5 antibody positivity (HR = 2.08, 95% CI = 1.64-13.22, $P = 0.032$) and CD3⁺CD8⁺ cell count lower than 31.38 cell/ μ L (HR = 8.6, 95% CI = 2.12-31.44, $P = 0.002$)

$P < 0.05$ are in bold

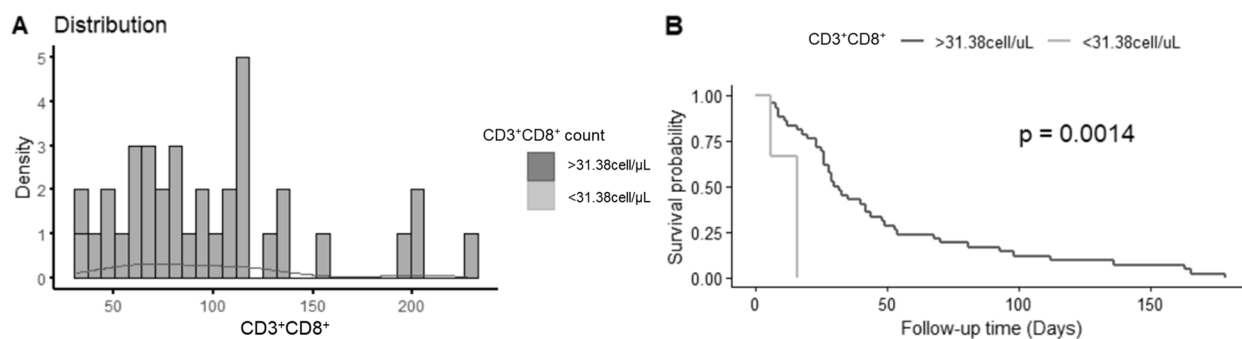


Fig. 4 **A, B** Optimal cut-off value (31.38 cell/ μ L) and the Kaplan–Meier survival curves for $CD3^+CD8^+$ count using the “*survminer*” R package. Figure shows the association between $CD3^+CD8^+$ cell count and survival outcome in patients with DM-ILD. Figure 3A is a histogram that shows the frequency of patients with recorded outcomes in two groups: those with $CD3^+CD8^+$ cell count lower than 31.38 cell/ μ L ($n=2$) and those with $CD3^+CD8^+$ cell count higher than 31.38 cell/ μ L ($n=43$). Figure 3B is a Kaplan–Meier curve that shows the survival probability of the two groups over time. The log-rank test revealed a significant difference in survival between the two groups ($P=0.0014$), with the higher $CD3^+CD8^+$ cell count group having a better prognosis than the lower $CD3^+CD8^+$ cell count group

[18, 28, 29], we focused on $MDA5^+$ DM-ILD by dividing DM-ILD into two groups based on $MDA5$ positivity in an attempt to obtain more pertinent clinical outcome indicators. By using $MDA5^-$ DM-ILD as controls, it will be more conducive to elucidating the characteristics of lymphocyte subsets in the $MDA5^+$ DM-ILD.

Data was collected prior to the treatment, therefore the influence of immunosuppressive drugs on lymphocyte subsets could be avoided. More DAD and RPILD were observed in the $MDA5^+$ DM-ILD after patients’ enrollment, while more NSIP was observed in the control group. Follow up data showed a mortality rate of 50.8% in $MDA5^+$ DM-ILD group and 7.7% in $MDA5^-$ DM-ILD group during the 180-day length (Table 1), further confirmed the much worse prognosis of $MDA5^+$ DM-ILD.

Compared to the $MDA5^-$ DM-ILD group, the $MDA5^+$ DM-ILD group exhibited significantly lower lymphocyte count, T lymphocyte count and B lymphocyte count. It was interesting to find that T and B lymphocytes decreased in an asynchronous fashion. T lymphocyte count was approximately 38.9% lower in $MDA5^+$ DM-ILD than in $MDA5^-$ DM-ILD, while B lymphocyte count was about 27.8% lower (Table 3). Data from Supplementary material also showed similar results when comparing death subjects to survival subjects. These results indicate that T lymphocytes might participate more in the pathogenesis. Further analysis on the lymphocyte subsets revealed that both $CD3^+CD4^+$ cell count and $CD3^+CD8^+$ cell count decreased, and their degrees of decrease were similar to that of total T lymphocytes. The mechanism underlying the lymphocytes decrease is largely unknown. Prior research has shown that the low T lymphocyte count in the peripheral blood of patients with DM can be attributed, in part, to the inhibited autophagy function of T cells, which promotes T

lymphocyte apoptosis [30]. Furthermore, massive immune cell infiltration was identified in lung tissue of $MDA5^+$ DM patients with ILD complications [31], it was hypothesized that activated lymphocytes in circulation were recruited to the target organs, such as lung [32]. The hypothesis was further supported by the increase of $CD3^+CD4^+$ count in alveolar lavage fluid of patients with DM [33]. Lymphocytes in peripheral blood metastasizing to the lungs where lymphatic vessels are abundant, participate in the local immune response and result in lymphopenia in peripheral blood. Similar results was found in B lymphocytes from tissues like muscle and lung biopsies [34, 35]. Future observations on the alterations of alveolar lavage fluid, lung biopsy tissue, and peripheral blood lymphocytes tracking in patients with $MDA5^+$ DM-ILD and $MDA5^-$ DM-ILD may provide additional support for the hypothesis.

Previous studies indicates that NK cells can release an excessive amount of $IFN-\gamma$, leading to pulmonary affection [36], and the total number of NK cells in myositis patients with pulmonary affection is lower than in those without pulmonary affection [37]. However, the count of peripheral blood NK cells did not differ significantly between the $MDA5^+$ DM-ILD and $MDA5^-$ DM-ILD groups in our study. The percentage of NK cells was statistically higher in the $MDA5^+$ DM-ILD group, which might be explained by drastically decreased total number of lymphocytes. When comparing the death subjects to the survival subjects, the percentage of NK cells did not differ, but the cell count was significantly reduced. Further research is warranted to clarify whether NK cells play a role in determining the clinical course.

The regression analysis in $MDA5^+$ DM-ILD patients showed that the poor prognosis was associated with low

CD3⁺CD8⁺ and low CD3⁺CD19⁺ levels (HR were 3.81 and 3.43, respectively) (Table 5). When the analysis was performed in all DM-ILD, low CD3⁺CD8⁺ cell count was independent predictor of death (HR 8.6) even after adjusting for anti-MDA5 and other clinical characteristics, and the HR was much higher than that of anti-MDA5 (HR 2.08) (Table 6). So, a low CD3⁺CD8⁺ might be a better prognostic factor than anti-MDA5 and warrant further study.

Limitation of the study

This study has several limitations. Firstly, the high prevalence of MDA5⁺ DM was unavoidable given that all of the clinical records collected for this retrospective study originated from grade A tertiary hospitals. Patient cohorts may represent a spectrum of more severe diseases due to referral bias. Secondly, all the participants were of East Asian descent, no other races were included. Thirdly, we did not continuously monitor post-treatment changes in lymphocyte subsets, which may have been associated with the treatment response. In the future, larger population-based multicenter studies will be required to obtain more accurate data.

Conclusions

Patients with MDA5⁺ DM-ILD exhibited significant immune imbalance characterized primarily by diminished T and B lymphocytes. Peripheral blood lymphocyte subsets may serve as prognostic markers for MDA5⁺ DM-ILD and DM-ILD. Moreover, lower CD3⁺CD8⁺ is an independent risk factor for the prognosis of MDA5⁺ DM-ILD and DM-ILD, laying a foundation for further prognostic prediction and targeted therapy.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12890-023-02706-y>.

Additional file 1: Supplementary material 1. Comparison of clinical data between survived and deceased patients in the MDA5⁺ DM-ILD cohort.

Acknowledgements

Not applicable.

Authors' contributions

FPR and LF participated in the study design. FPR conducted the study and wrote the manuscript. TL, WCZ, LG, ZLZ and YWS collected and processed medical data. QC, SSY and SWG analyzed the data. DW, XYX, XLZ and XYB revised and edited the manuscript. FPR revised the final manuscript. All authors read and approved the final manuscript.

Funding

There is no funding allocated for this retrospective study.

Availability of data and materials

The data that support the findings of this study are not openly available due to reasons of sensitivity and are available from the corresponding author upon reasonable request.

Declarations

Ethics approval and consent to participate

The study was conducted in accordance with the Declaration of Helsinki, and the protocol was approved by the Ethics Committee of Chao-Yang Hospital, Capital Medical University (Project ID 2023-4-10-3), which waived the need for written informed consent because of the retrospective study.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Author details

¹Beijing Shijitan Hospital, Capital Medical University, Beijing, China. ²University of Pittsburgh, Pittsburgh, USA. ³Beijing Chao-Yang Hospital, Capital Medical University, Beijing, China. ⁴PLA Strategic Support Force Medical Center, Beijing, China. ⁵Xuanwu Hospital, Capital Medical University, Beijing, China. ⁶Weifang Medical College, Weifang, China.

Received: 19 June 2023 Accepted: 11 October 2023

Published online: 28 October 2023

References

- Sugiyama Y, Yoshimi R, Tamura M, et al. The predictive prognostic factors for polymyositis/dermatomyositis-associated interstitial lung disease. *Arthritis Res Ther*. 2018;20(1):7.
- Zhang H, Yue J, Hou X, et al. Rapidly progressive interstitial lung disease combined with pneumocystis jiroveci pneumonia in a patient with single anti-TIF-1 γ antibody positive dermatomyositis in the context of an underlying tumor. *BMC Pulm Med*. 2023;23(1):248.
- Betteridge Z, McHugh N. Myositis-specific autoantibodies: an important tool to support diagnosis of myositis. *J Intern Med*. 2016;280(1):8–23.
- Kochi Y, Kamatani Y, Kondo Y, et al. Splicing variant of WDFY4 augments MDA5 signalling and the risk of clinically amyopathic dermatomyositis. *Ann Rheum Dis*. 2018;77(4):602–11.
- Alenzi FM. Myositis Specific Autoantibodies: A Clinical Perspective. *Open Access Rheumatol*. 2020;12:9–14.
- Wen L, Chen X, Cheng Q, et al. Myositis-specific autoantibodies and their clinical associations in idiopathic inflammatory myopathies: results from a cohort from China. *Clin Rheumatol*. 2022;41(11):3419–27.
- Motegi SI, Sekiguchi A, Toki S, et al. Clinical features and poor prognostic factors of anti-melanoma differentiation-associated gene 5 antibody-positive dermatomyositis with rapid progressive interstitial lung disease. *Eur J Dermatol*. 2019;29(5):511–7.
- Allenbach Y, Uzunhan Y, Toquet S, et al. Different phenotypes in dermatomyositis associated with anti-MDA5 antibody: Study of 121 cases. *Neurology*. 2020;95(1):e70–8.
- Wu W, Guo L, Fu Y, et al. Interstitial lung disease in Anti-MDA5 positive dermatomyositis. *Clin Rev Allergy Immunol*. 2021;60(2):293–304.
- Moghadam-Kia S, Oddis CV, Sato S, Kuwana M, Aggarwal R. Antimelanoma differentiation-associated Gene 5 antibody: expanding the clinical spectrum in North American patients with dermatomyositis. *J Rheumatol*. 2017;44(3):319–25.
- Temmoku J, Sato S, Fujita Y, et al. Clinical significance of myositis-specific autoantibody profiles in Japanese patients with polymyositis/dermatomyositis. *Medicine (Baltimore)*. 2019;98(20): e15578.
- DeWane ME, Waldman R, Lu J. Dermatomyositis: Clinical features and pathogenesis. *J Am Acad Dermatol*. 2020;82(2):267–81.
- Ceribelli A, De Santis M, Isailovic N, Gershwin ME, Selmi C. The immune response and the pathogenesis of idiopathic inflammatory myositis: a critical review. *Clin Rev Allergy Immunol*. 2017;52(1):58–70.

14. Hervier B, Uzunhan Y. Inflammatory myopathy-related interstitial lung disease: from pathophysiology to treatment. *Front Med (Lausanne)*. 2019;6:326.
15. Cassius C, Amode R, Delord M, et al. MDA5(+) dermatomyositis is associated with stronger skin type I interferon transcriptomic signature with upregulation of IFN- κ transcript. *J Invest Dermatol*. 2020;140(6):1276-1279.e7.
16. Huang W, Ren F, Luo L, et al. The characteristics of lymphocytes in patients positive for anti-MDA5 antibodies in interstitial lung disease. *Rheumatology (Oxford)*. 2020;59(12):3886–91.
17. Chen F, Wang D, Shu X, Nakashima R, Wang G. Anti-MDA5 antibody is associated with A/SIP and decreased T cells in peripheral blood and predicts poor prognosis of ILD in Chinese patients with dermatomyositis. *Rheumatol Int*. 2012;32(12):3909–15.
18. Li W, Tian X, Lu X, et al. Significant decrease in peripheral regulatory B cells is an immunopathogenic feature of dermatomyositis. *Sci Rep*. 2016;6:27479.
19. Selva-O'Callaghan A, Romero-Bueno F, Trallero-Araguás E, et al. Pharmacologic treatment of Anti-MDA5 rapidly progressive interstitial lung disease. *Curr Treatm Opt Rheumatol*. 2021;7(4):319–33.
20. Mao MM, Xia S, Guo BP, et al. Ultra-low dose rituximab as add-on therapy in anti-MDA5-positive patients with polymyositis /dermatomyositis associated ILD. *Respir Med*. 2020;172: 105983.
21. Ge Y, Li S, Tian X, He L, Lu X, Wang G. Anti-melanoma differentiation-associated gene 5 (MDA5) antibody-positive dermatomyositis responds to rituximab therapy. *Clin Rheumatol*. 2021;40(6):2311–7.
22. Bohan A, Peter JB. Polymyositis and dermatomyositis (first of two parts). *N Engl J Med*. 1975;292(7):344–7.
23. Bohan A, Peter JB. Polymyositis and dermatomyositis (second of two parts). *N Engl J Med*. 1975;292(8):403–7.
24. Sontheimer RD. Cutaneous features of classic dermatomyositis and amyopathic dermatomyositis. *Curr Opin Rheumatol*. 1999;11(6):475–82.
25. Chawla A, Kumar T, Mukherjee P. Antisynthetase syndrome: Initial and follow-up imaging features. *Lung India*. 2018;35(6):523–5.
26. Lian X, Zou J, Guo Q, et al. Mortality risk prediction in amyopathic dermatomyositis associated with interstitial lung disease: the FLAIR model. *Chest*. 2020;158(4):1535–45.
27. Li Y, Li Y, Wu J, et al. Predictors of poor outcome of Anti-MDA5-Associated rapidly progressive interstitial lung disease in a Chinese cohort with dermatomyositis. *J Immunol Res*. 2020;2020:2024869.
28. Wang DX, Lu X, Zu N, et al. Clinical significance of peripheral blood lymphocyte subsets in patients with polymyositis and dermatomyositis. *Clin Rheumatol*. 2012;31(12):1691–7.
29. Feng M, Guo H, Zhang C, et al. Absolute reduction of regulatory T cells and regulatory effect of short-term and low-dose IL-2 in polymyositis or dermatomyositis. *Int Immunopharmacol*. 2019;77: 105912.
30. Shu X, Chen F, Peng Q, et al. Potential role of autophagy in T-cell survival in polymyositis and dermatomyositis. *Mol Med Rep*. 2017;16(2):1180–8.
31. Ye Y, Chen Z, Jiang S, et al. Single-cell profiling reveals distinct adaptive immune hallmarks in MDA5+ dermatomyositis with therapeutic implications. *Nat Commun*. 2022;13(1):6458.
32. Mukae H, Ishimoto H, Sakamoto N, et al. Clinical differences between interstitial lung disease associated with clinically amyopathic dermatomyositis and classic dermatomyositis. *Chest*. 2009;136(5):1341–7.
33. Galindo-Feria AS, Albrecht I, Fernandes-Cerqueira C, et al. Proinflammatory Histidyl-Transfer RNA Synthetase-Specific CD4+ T Cells in the Blood and Lungs of Patients With Idiopathic Inflammatory Myopathies. *Arthritis Rheumatol*. 2020;72(1):179–91.
34. Dzangué-Tchoupou G, Allenbach Y, Preuße C, Stenzel W, Benveniste O. Mass cytometry reveals an impairment of B cell homeostasis in anti-synthetase syndrome. *J Neuroimmunol*. 2019;332:212–5.
35. Yamadori I, Fujita J, Kajitani H, et al. Lymphocyte subsets in lung tissues of interstitial pneumonia associated with untreated polymyositis/dermatomyositis. *Rheumatol Int*. 2001;21(3):89–93.
36. Noyola DE, Juárez-Vega G, Monjarás-Ávila C, et al. NK cell immunophenotypic and genotypic analysis of infants with severe respiratory syncytial virus infection. *Microbiol Immunol*. 2015;59(7):389–97.
37. Pawlitzki M, Nelke C, Rolfes L, et al. NK Cell patterns in idiopathic inflammatory myopathies with pulmonary affection. *Cells*. 2021;10(10):2551.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

