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Analysis of clinical characteristics and prognosis of lung cancer patients with CPFE or COPD: a retrospective study



Yuying Wei¹, Liuqing Yang¹ and Qing Wang^{1*}

Abstract

Background Lung cancer (LC) commonly occurs in patients with combined pulmonary fibrosis and emphysema (CPFE) and chronic obstructive pulmonary disease (COPD), but comparative research is limited. This study examines clinical characteristics, treatments, and prognosis in LC patients with CPFE or COPD.

Methods The retrospective study involved 75 lung cancer patients with CPFE and 182 with COPD. It analyzed clinical features, tumor pathology, pulmonary function, laboratory parameters, and treatment responses.

Results Notable differences were found between the CPFE+LC and COPD+LC groups. Both groups were mostly elderly, male smokers. The CPFE+LC group had higher BMI and more adenocarcinoma and squamous cell carcinoma, while COPD+LC had predominantly squamous cell carcinoma. CPFE+LC tumors were mostly in the lower lobes; COPD+LC's were in the upper lobes. The CPFE+LC group showed higher tumor metastasis rates, more paraseptal emphysema, and elevated levels of TG, CEA, NSE, and Killer T Cells. In advanced stages (IIIB-IV), the CPFE+LC group receiving first-line treatment had shorter median progression-free survival (PFS) and a higher risk of progression or death than the COPD+LC group, regardless of whether it was non-small cell lung cancer (NSCLC) or small cell lung cancer (SCLC). No significant PFS difference was found within CPFE+LC between chemotherapy and immunotherapy, nor in immune-related adverse events between groups, with interstitial pneumonia being common.

Conclusion This study emphasizes distinct lung cancer characteristics in CPFE or COPD patients, highlighting the need for tailored diagnostic and treatment approaches. It advocates for further research to improve care for this high-risk group.

Keywords Lung cancer (LC), Combined pulmonary fibrosis and emphysema (CPFE), Chronic obstructive pulmonary disease (COPD), Progression-free survival (PFS)

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Introduction

Lung cancer (LC) is a global healthcare concern, representing the most prevalent form of cancer worldwide, accounting for 11.6% of all cancer cases and standing as the leading cause of cancer-related fatalities. In 2018, over 1.7 million lives were claimed by lung cancer [1, 2].

Chronic obstructive pulmonary disease (COPD) is associated with a high disease burden, and according to predictions by the World Health Organization, it will become the third leading cause of death by 2030 [3]. Both lung cancer and COPD are highly associated with smoking, and COPD is an independent risk factor for the development of lung cancer [4]. Reports indicate that COPD affects a significant proportion of lung cancer patients worldwide, ranging from 45 to 63% [5].

Combined pulmonary fibrosis and emphysema (CPFE) is a unique clinical entity [6], with a prevalence of 26–54% among patients with idiopathic pulmonary fibrosis (IPF) [7–9]. According to an Official ATS/ERS/JRS/ALAT Clinical Practice Guideline, approximately 2–52% of CPFE patients eventually develop lung cancer [7]. High-resolution computed tomography (HRCT) shows emphysema in the upper lobes of the lung along with fibrosis in the lower lobes [6]. CPFE often presents with impaired gas exchange, and is prone to complications such as pulmonary hypertension and lung cancer [6, 10, 11].

Both COPD and CPFE represent chronic lung diseases that are frequently encountered among elderly male smokers [7]. They share radiological evidence of pulmonary emphysema, and the incidence of concurrent lung cancer is significantly elevated in both conditions. However, patients with CPFE or COPD demonstrate notable differences in terms of pathology, physiology, clinical presentation, radiology, and prognosis [12]. Currently, research comparing lung cancer patients with CPFE or COPD remains limited, underscoring the need for more extensive exploration.

This study aims to delve into the clinical characteristics and treatment outcomes of lung cancer patients with CPFE or COPD using a retrospective approach based on real-world data. This research not only helps in comprehensively understanding these two complex diseases but also holds significant importance in developing precision medicine strategies for these high-risk groups.

Methods

Study design and patient selection

This retrospective study included 75 lung cancer patients with CPFE (CPFE+LC group) and 182 lung cancer patients with COPD (COPD+LC group) who were admitted to the Department of Respiratory and Critical Care Medicine at the First Affiliated Hospital, Zhejiang University School of Medicine. The patients were selected consecutively from January 2021 to December 2022. All participants were aged 60 years or older. The follow-up date was until June 2023. Our study was conducted in compliance with ethical standards for research involving human subjects. The Ethics Board of the First Affiliated Hospital of Zhejiang University approved this study.

Inclusion criteria

- (1) CPFE: Diagnosis was based on Cottin's 2005 criteria
 [6], HRCT scans demonstrate emphysematous changes predominantly distributed in the upper lungs, characterized by the presence of low attenuation areas with thin walls (<1 mm) and no clear boundaries, or multiple pulmonary bullae with diameters > 1 cm. The extent of pulmonary emphysema within the lung fields should be ≥ 10.0%. Additionally, HRCT of the chest should show fibrotic changes primarily involving the lower lungs and subpleural areas, characterized by a reticular pattern, as well as varying degrees of honeycombing and/or traction bronchiectasis.
- (2) COPD: Based on the GOLD 2023 guidelines [13], patients had a discharge diagnosis of COPD, with CT scans revealing increased and thickened pulmonary markings, bilateral lung fields with areas of low attenuation characterized by the absence of walls or extremely thin walls, or the presence of pulmonary bullae (diameter ≥ 1 cm, wall thickness ≤ 1 mm).
- (3) Lung cancer: Confirmation of tumor cells through biopsy, surgical pathology, or cytological examination, including lymph node biopsy, sputum cytology, or pleural fluid cytology. Patients exhibited an Eastern Cooperative Oncology Group (ECOG) performance status ranging from 0 to 2.

Exclusion criteria

- (1) Excluding conditions such as nodular diseases, granulomatosis with polyangiitis, allergic alveolitis, lymphangioleiomyomatosis, eosinophilic pneumonia, Langerhans cell histiocytosis, pulmonary alveolar proteinosis, idiopathic pulmonary hemosiderosis, drug- or treatment-related interstitial changes (e.g., pesticides, radiation).
- (2) Excluding patients with lung metastases from other tumors, severe liver or kidney dysfunction, hematological malignancies, severe cardiovascular or cerebrovascular diseases, concurrent viral infections, asthma, or a large amount of pleural effusion/ pneumothorax.
- (3) Excluding patients with missing HRCT imaging and pulmonary function results during hospitalization,

those whose lung cancer pathology cannot be classified, and those with incomplete medical records and relevant examination data.

Data collection

The patient data was retrieved from the electronic medical records, including demographics, laboratory results, tumor pathology, TNM staging (the eighth edition) [14], tumor location, comorbidities, HRCT images, and pulmonary function test results. Document information of advanced cancer patients with TNM staging ranging from IIIB to IV. Take note of whether these patients underwent immunotherapy, record immunotherapyrelated adverse reactions (AEs), document the initiation date of first-line and second-line chemotherapy, and make a record of the time of the first occurrence of disease progression (switch to second-line chemotherapy or imaging evidence of tumor progression) or death (whichever occurred first).

Statistical analysis

Data analysis was conducted using SPSS version 25.0. Graphing was performed using GraphPad Prism 8. Categorical variables were summarized as frequencies and percentages, and continuous variables were described using mean±standard deviation or median with interquartile range. We used the Mann-Whitney U test for continuous variables, and the Chi-squared(χ^2) test or Fisher's exact test for categorical data, with Bonferroni correction for multiple comparisons. The Log-rank test was used to compare survival differences between CPFE+LC group and COPD+LC group, and the Kaplan-Meier method was used to construct survival curves for progression-free survival (PFS). *P*<0.05 was considered statistically significant.

Results

Demographics and clinical comorbidities

As shown in Table 1, the CPFE+LC group consisted exclusively of male patients (100%, n=75), while the COPD+LC group was predominantly male (99.45%, n=182). The median age for both groups was 69 years. A significant disparity was observed in BMI, with the CPFE+LC group showing a higher average BMI (22.85±3.43) compared to the COPD+LC group (21.51±2.94), p=0.002.

The prevalence of smoking was high in both groups, with all the CPFE+LC patients and 96.70% of the COPD+LC patients being current or former smokers. No significant difference in smoking amount (pack-years) was observed. Comorbid conditions such as bronchiectasis, pulmonary arterial hypertension, connective tissue diseases, coronary artery atherosclerosis, hypertension, diabetes, prior pulmonary tuberculosis, pulmonary embolism and peripheral vascular disease had similar rates in both groups. However, the incidence of

 Table 1
 Clinical characteristics and comorbidities in CPFE+LC and COPD+LC patients

Variable	CPFE + LC (n = 75)	COPD + LC (n = 182)	Statistic	Р
Gender (men)	75 (100.00%)	181 (99.45%)	-	1.000
Age	69 (66,74)	69 (66,73)	Z=-0.413	0.679
BMI (kg/m²)	22.85 ± 3.43	21.51 ± 2.94	t=-3.147	0.002
ECOG (score)			χ ² =3.828	0.148
0	46 (61.30%)	121 (66.50%)		
1	25 (33.30%)	59 (32.40%)		
2	4 (5.30%)	2 (1.10%)		
Ex- or current smokers	75 (100.00%)	176 (96.70%)	χ ² =1.292	0.256
Smoking amount (pack-years)	40.00 (30.00-60.00)	40.00 (30.00-60.00)	Z=0.428	0.671
Bronchiectasia	3 (4.00%)	18 (9.89%)	χ ² =2.456	0.117
Pulmonary hypertension	12 (16.00%)	26 (14.29%)	χ ² =0.124	0.725
Connective tissue disease	2 (2.67%)	4 (2.20%)	χ ² =0.000	1.000
Coronary artery atherosclerosis	43 (57.33%)	89 (48.90%)	$\chi^2 = 1.512$	0.219
Hypertension	31 (41.33%)	71 (39.01%)	χ ² =0.120	0.729
Diabetes mellitus	8 (10.67%)	23 (12.64%)	χ ² =0.194	0.659
Previous pulmonary tuberculosis	1 (1.33%)	8 (4.40%)	χ ² =0.707	0.400
Previous cerebral infarction	6 (8.00%)	1 (0.50%)	χ ² =8.493	0.004
Pulmonary embolism	3 (4.00%)	4 (2.20%)	χ ² =0.149	0.700
Peripheral vascular disease	6 (8.00%)	13 (7.10%)	χ ² =0.057	0.811
Dust exposure	0 (0.00)	2 (1.10%)	-	1.000
Family history of cancer	7 (9.33%)	15 (8.24%)	χ ² =0.081	0.776

BMI: body mass index; ECOG: Eastern Cooperative Oncology Group

previous cerebral infarction was significantly higher in the CPFE+LC group, p=0.004.

Lung cancer pathology and radiological findings

There was a statistically significant difference in the pathology of lung cancer between the two groups (p=0.004). Adenocarcinoma (36.00%) and squamous cell carcinoma (34.67%) were more common in CPFE, while squamous cell carcinoma (52.75%) was more common in COPD. The distribution of tumors differed between the two groups (p < 0.0167), with a higher incidence of lung cancer in the lower lobes (60.00%) in the CPFE+LC group compared to the COPD+LC group (40.66%). The CPFE+LC group also had a higher metastasis rate (49.33%) than the COPD+LC group (32.97%) (p=0.014). It's worth noting that nearly half of the patients in the CPFE+LC group were diagnosed with stage IV lung cancer (49.33%), whereas in the COPD+LC group, there were more patients at stage III (44.51%), though this difference was not statistically significant. Statistical analysis revealed that the types of emphysema were significantly different between the two groups (p < 0.001). Both panlobular emphysema and paraseptal emphysema exhibited intergroup differences in both groups (both p < 0.0167). Paraseptal emphysema was the most common type in the CPFE+LC group (41.33%), while centrilobular emphysema had the highest prevalence in the COPD+LC group (60.99%). The results are indicated in Table 2.

Pulmonary function test parameters

As shown in Table 2, pulmonary function parameters such as FEV1 and FEV1/ FVC were higher in the CPFE+LC group (all p<0.001). In contrast, DLCO (p<0.001), DLCO/ VA (p=0.009), RV (p<0.001), and RV/ TLC (p<0.001) were significantly lower in the CPFE+LC group compared to the COPD+LC group.

Laboratory test results

As shown in Table 3, the CPFE+LC group exhibited significantly higher levels of TG (p=0.040), CEA (p=0.010), NSE (p=0.003), and killer T cell counts (CD3+, CD8+) (p=0.028) compared to the COPD+LC group. But there were no statistically significant differences in complete blood count, immunoglobulins, complement, and inflammatory cytokines.

We have compiled the PFS data of the first-line treatment for all advanced cancer patients with TNM staging ranging from IIIB to IV. The results indicate that in the CPFE+LC group, comprising 51 patients, the median PFS of first-line treatment was 6.0 months (95% CI: 4.9–7.1). In contrast, the COPD+LC group, consisting of 99 patients, exhibited a longer median PFS of 9.0 months (95% CI: 6.2–11.8). Statistical analysis revealed a significant difference in PFS between the two groups (p=0.0003, χ^2 =13.29). Comparing the CPFE+LC group

to the COPD+LC group, the former showed an 85.7% increased risk of disease progression or death after receiving first-line chemotherapy, with a hazard ratio (HR) of 1.857 (95% CI: 1.239–2.783). Figure 1 illustrates Kaplan- Meier survival curves for two groups undergoing pharmacological treatment.

In all advanced lung cancer patients with TNM staging ranging from IIIB to IV who received first-line treatment, we conducted subgroup survival analyses separately for patients with non-small cell lung cancer (NSCLC) and small cell lung cancer (SCLC). In the CPFE+LC group, consisting of 33 NSCLC patients, the median PFS was 6.0 months (95% CI: 4.491-7.509). In the COPD+LC group, comprising 80 NSCLC patients, the median PFS was 9.0 months (95% CI: 6.597-11.403). Statistical analysis revealed a significant difference in PFS between the two groups (p=0.0110, $\chi^2=6.464$). After receiving firstline treatment, the CPFE+LC group showed a 69.3% increased risk of disease progression or death, with a HR of 1.693 (95% CI: 1.035-2.770). Figure 2 displays the Kaplan- Meier survival curves for the two groups of NSCLC patients following pharmacological treatment.

In the CPFE+LC group, consisting of 18 SCLC patients, the median PFS was 5.5 months (95% CI: 3.337–6.663). In the COPD+LC group, comprising 19 SCLC patients, the median PFS was 11.0 months (95% CI: 6.782–15.218). Statistical analysis indicated a significant difference in PFS between the two groups (p=0.0173, χ^2 =5.667). Following first-line treatment, the CPFE+LC group exhibited a 109.1% increased risk of disease progression or death, with a HR of 2.091 (95% CI: 1.029–4.247). Figure 3 displays the Kaplan- Meier survival curves for the two groups of SCLC patients receiving pharmacological treatment.

We further analyzed the 51 patients in the CPFE+LC group mentioned above. Among them, 23 patients received chemotherapy as first-line treatment, with a median PFS of 6.0 months (95% CI: 4.9–7.1). The remaining 28 patients received a combined treatment with immunotherapy and chemotherapy, and their median PFS of first-line treatment was 6.0 months (95% CI: 4.3–7.7), with an HR of 0.672 (95% CI: 0.373–1.211) (p=0.119, χ^2 =2.42). These results indicate that, in the CPFE+LC group, there was no significant difference in the median PFS of first-line treatment between the chemotherapy-only group and the combined treatment group. Figure 4 illustrates the Kaplan- Meier survival curves for two treatment groups within the CPFE+LC group.

Among the CPFE+LC group, consisting of 28 patients who received immunotherapy, 10 cases (35.7%) reported immune-related AEs. In contrast, among the 78 lung cancer patients with COPD who were treated with immunotherapy, 19 cases (24.4%) experienced immune-related

Table 2	Comparison c	of tumor characteristics,	radiographic	distribution,	, and pulmonary	/ function	between lu	ng cancer	patients with
CPFE or C	OPD								

Variable	CPFE+LC (n = 75)	COPD + LC (n = 182)	Statistic	Р
Pathological classification			χ ² =13.100	0.004
Adenocarcinoma	27 (36.00%)	64 (35.16%)		
Squamous cell carcinoma	26 (34.67%)	96 (52.75%)		
Small cell carcinoma	18 (24.00%)	19 (10.44%)		
Other	4 (5.33%)	3 (1.65%)		
Distribution of left/ right lung lobes			χ ² =0.750	0.386
Right lung	44 (58.67%)	96 (52.75%)		
Left lung	31 (41.33%)	86 (47.25%)		
Distribution of lung lobe location			χ ² =8.557	0.014
Upper lobe	26 (34.67%) *	99 (54.40%)		
Middle lobe	4 (5.33%)	9 (4.95%)		
Lower lobe	45 (60.00%) *	74 (40.66%)		
Т			χ ² =2.545	0.467
1	9 (12.00%)	28 (15.38%)		
2	24 (32.00%)	62 (34.07%)		
3	13 (17.33%)	19 (10.44%)		
4	29 (38.67%)	73 (40.11%)		
Ν			$\chi^2 = 5.060$	0.167
0	14 (18.67%)	34 (18.68%)		
1	5 (6.67%)	27 (14.84%)		
2	30 (40.00%)	77 (42.31%)		
3	26 (34.67%)	44 (24.18%)		
Μ			$\chi^2 = 6.054$	0.014
0	38 (50.67%)	122 (67.03%)	~	
1	37 (49.33%)	60 (32.97%)		
Stage			x ² =6.723	0.081
	3 (4.00%)	16 (8.79%)	~	
11	8 (10.67%)	25 (13.74%)		
111	27 (36.00%)	81 (44.51%)		
IV	37 (49.33%)	60 (32.97%)		
Types of emphysema			x ² =30.028	< 0.001
Panlobular emphysema	24 (32.00%)	44 (24.18%)	~	
Centrilobular emphysema	20 (26.67%) #	111 (60.99%)		
Paraseptal emphysema	31 (41.33%) #	27 (14.84%)		
FEV1 (L)	1.92 ± 0.43	1.59 ± 0.54	t=-5.169	< 0.001
FVC (L)	2.69+0.52	2.60+0.64	t=-1.276	0.204
FEV1/ FVC (%)	71.19 (64.65–76.89)	61.25 (52.25–66.49)	Z=7.206	< 0.001
VC (L)	2.30 (1.97–2.73)	2.21 (1.76–2.71)	Z=1.105	0.269
DI CO (ml /min/mmHa)	8.48 (4.02–11.59)	10.11 (4.35–14.81)	7=2.398	0.016
DLCO/VA (mL/min/mmHa/L)	2.02 (0.85–2.96)	2.69 (1.00–3.40)	Z=2.595	0.009
RV (L)	1.97 (1.60–2.41)	2.35 (1.92–2.81)	Z=3.945	< 0.001
TLC (L)	4.18 (3.70–4.84)	4.51 (3.74–5.20)	Z=1.671	0.095
RV/TIC (%)	47.20 (40.84–52.55)	52.11 (46.50–57.65)	Z=4.605	< 0.001

 *P < 0.0167 when compared to the COPD+LC group

 $^{\#}P$ < 0.0167 when compared to the COPD+LC group

T: tumor; N: node; M: metastasis; FEV1: forced expiratory volume in 1 s; FVC: forced vital capacity; FEV1/ FVC: forced expiratory volume in 1 s to forced vital capacity ratio; VC: vital capacity; DLCO: diffusing capacity of the lung for carbon monoxide; DLCO/ VA: diffusing capacity per unit of alveolar volume; RV: residual volume; TLC: total lung capacity; RV/ TLC: residual volume to total lung capacity ratio

Table 3 Laboratory test results in lung cancer patients with CPFE or COPD

WBC (x10 ¹ /L) 660 (539-870) 6.63 (572-784) 2=0.41 0.521 PLT (x10 ¹ /L) 215.00 (17250-22/5.00) 234.00 (18250-296.00) Z=1.632 0.103 ALC (x10 ¹ /L) 1.52 (1.19-1.86) 1.39 (1.02-1.78) Z=1.73 0.081 TG (mmoL/L) 3.93 (3.30-4.76) 4.17 (3.54-4.74) Z=2.053 0.040 TG (mmoL/L) 3.93 (3.30-4.76) 4.17 (3.54-4.74) Z=1.072 0.020 TD (mmoL/L) 2.07 (1.77-2.83) 2.31 (1.84-2.83) Z=1.131 0.270 FB (mq/L) 3.65 (2.87-4.93) 3.58 (2.96-4.78) Z=0.232 0.817 G (mg/dL) 1.20 2.22 8.13 119.28 ± 2.297 t=0.774 0.441 IgG (mg/dL) 1.30 0.22 ± 3.95 2.3 1.20 (1.21 ± 3.11.91 t=1.171 0.245 IgA (mg/L) 3.80 (2.00-7.62) 3.00 (1.90-570.01 Z=1.561 0.207 IgA (mg/L) 3.80 (2.00-7.62) 3.00 (1.90-6.70) Z=1.839 0.606 SE forg/mL) 1.20 (1.07-2.27) 1.20 (0.80-2.90) Z=1.839 0.606 SE forg/mL) 1.20 (1.52-2.4	Variable	CPFE+LC (<i>n</i> =75)	COPD + LC (<i>n</i> = 182)	Statistic	Р
PLT (k10 ² /l.) 21500 (17250-2750) 24400 (18825-29600) 2=1.632 0.103 ANC (x10 ² /l.) 4.40 (327-592) 4.38 (3.36-5.62) Z=0.256 0.078 ALC (x10 ² /l.) 1.52 (1.19-1.86) 1.39 (102-1.78) Z=1.743 0.081 TG (mmol./l.) 1.16 (0.87-1.60) 0.99 (0.75-1.43) Z=0.256 0.040 TC (mmol./l.) 3.93 (3.0-4.76) 4.17 (3.54-4.74) Z=1.272 0.203 LD Immol./l.) 2.07 (1.77-2.83) 2.31 (1.42-2.83) Z=1.103 0.270 CRP (mg/L) 8.60 (3.0-20.67) 4.77 (2.45-2.350) Z=1.33 0.877 Fibrinogen (g/L) 3.58 (2.96-4.78) Z=0.323 0.877 IG (mg/dL) 170.23 ± 28.13 119.28 ± 2.377 I=-0.323 0.747 IgG (mg/dL) 170.23 ± 28.13 119.28 ± 2.377 I=-0.323 0.747 IgG (mg/dL) 7.900 (10.00-310.00) 95.10 (56.25-217.25) Z=1.276 0.202 IgA (mg/LL) 7.900 (10.00-97.00) 92.10 (59.25-127.25) Z=1.276 0.203 IgA (mg/LL) 1.20 (0.09-7.00) 92.10 (59.25-127.25) Z=1.276 0.204 IgA (mg/LL	WBC (×10 ⁹ /L)	6.60 (5.39–8.70)	6.63 (5.72–7.84)	Z=0.641	0.521
AIC (x10 ² /L) 4.40 (327-5.92) 4.38 (3.38-5.62) Z=0.256 0.798 ALC (x10 ² /L) 1.52 (1.19-1.86) 1.39 (1.02-1.78) Z=1.743 0.081 TC (mmoL/L) 1.16 (0.87-1.60) 0.99 (0.27-1.43) Z=2.053 0.040 TC (mmoL/L) 2.07 (1.77-2.83) 2.31 (1.84-2.83) Z=1.103 0.270 CRP Img/L) 3.67 (2.87-4.93) 3.58 (2.96-4.78) Z=0.222 0.817 CA (mg/dL) 2.765.2.795 2.947 4.255 Z=0.723 0.747 CA (mg/dL) 1.02.32 ±.28.13 119.28 ±.22.97 t=0.323 0.747 IgG (mg/dL) 1.02.32 ±.395.23 1.01.21 ±.11.191 t=1.117 0.242 IgM (mg/dL) 2.81.00 (190.00-310.00) 1.95.50 (156.75-283.50) Z=1.561 0.118 CFA (ng/mL) 6.20 (3.50-9.95) 4.30 (3.00-747) Z=2.591 0.010 Viffa 21 -1 Ing/mL) 3.80 (2.0-76.2) 3.00 (10-96.70) Z=1.839 0.066 NSE (ng/mL) 1.840 (13.35-26.48) 1.560 (12.83-20.00) Z=0.297 0.003 SC (ng/mL) 1.20 (1.0-2.27) 1.20 (0.80-2.00) Z=0.297 0.003	PLT (×10 ⁹ /L)	215.00 (172.50–275.00)	234.00 (188.25–296.00)	Z=1.632	0.103
ALC (x10 ⁷ /L) 1.52 (1.19-1.86) 1.39 (1.02-1.78) Z = 1.743 0.081 TG (mmoL/L) 1.16 (0.87-1.60) 0.99 (0.75-1.43) Z = 2.053 0.040 TC (mmoL/L) 3.93 (3.02-476) 4.17 (3.54-4.74) Z = 1.103 0.270 DL (mmoL/L) 3.60 (3.02-0.667) 4.77 (2.45-2.35) Z = 1.319 0.187 Fibrinogen (xPL) 3.67 (2.87-4.93) 3.58 (2.96-4.78) Z = 0.323 0.747 UgG (mg/dL) 12.132 ± 2.8.13 119.28 ± 2.297 t= 0.323 0.747 UgG (mg/dL) 130.32 ± 395.23 1201 (2.1 ± ±1.191 t=t-1.11 0.245 UgM (mg/dL) 7.900 (51.00-9700) 92.10 (59.25-127.25) Z = 1.276 0.202 UgA (mg/dL) 2.810 (190.00-310.00) 19.55 0 (15.675-283.50) Z = 1.561 0.118 C KA (ng/mL) 3.80 (2.02-7.62) 3.00 (190-670) Z = 1.839 0.066 NSE (ng/mL) 1.20 (100-2.27) 1.20 (0.82-2.00) Z = 0.926 0.339 U (2.100/mL) 1.20 (0.7-1.83) 1.23 (0.36-2.34) Z = 1.490 0.137 U (2.100/mL) 1.20 (0.7-1.82) 2.72 (0.89-5.03) Z = 1.442 0.1137<	ANC (×10 ⁹ /L)	4.40 (3.27–5.92)	4.38 (3.38–5.62)	Z=0.256	0.798
TG (mmol/L) 1.16 (0.87–16.0) 0.99 (0.75–1.4) Z=1053 0.040 TC (mmol/L) 3.93 (3.0–4.76) 4.17 (3.54–4.74) Z=122 0.020 DL (mmol/L) 2.07 (1.77–2.83) 3.51 (1.84–2.83) Z=11.03 0.270 C4P (mg/L) 8.60 (3.20–2.067) 4.77 (2.45–2.35.0) Z=1.319 0.187 Flbrinogen (g/L) 3.56 (2.87–4.93) 3.58 (2.96–4.78) Z=0.0232 0.747 C4 (mg/dL) 121.32±2.81.3 119.28±2.297 t=0.323 0.747 IgG (mg/dL) 130.32 ± 395.23 1201.21 ± 311.91 t=1.171 0.245 IgA (mg/dL) 281.00 (190.00–310.00) 195.50 (156.75–283.50) Z=1.561 0.118 IgA (mg/dL) 281.00 (190.00–310.00) 195.50 (156.75–283.50) Z=1.561 0.118 SE (ng/mL) 1.80 (13.35–26.48) 15.60 (12.83–2000) Z=0.589 0.066 SE (ng/mL) 1.80 (13.35–26.48) 15.60 (12.83–2000) Z=0.589 0.0379 SE (ng/mL) 1.814 (01.35–26.472) 2.00 (0.80–2.00) Z=0.589 0.0379 SE (ng/mL) 1.82 (0.76–2.479) 7.96 (4.33–2.281) 0.018 0.1379	ALC (×10 ⁹ /L)	1.52 (1.19–1.86)	1.39 (1.02–1.78)	Z=1.743	0.081
TC (mmoL/L) 3.93 (3.30–4.76) 4.17 (3.54–4.74) Z=1222 0.203 LD (mmoL/L) 2.07 (1.77–2.83) 2.31 (1.84–2.83) Z=1.103 0.0270 FIP (mg/L) 8.60 (3.20–20.67) 47.77 (2.45–2.350) Z=1.023 0.817 Fibrinogen (g/L) 2.66 ±7.95 9.947 ±9.25 t=0.074 0.414 G3 (mg/dL) 121.32 ± 28.13 119.28 ± 2.97 t=0.323 0.747 IgA (mg/dL) 1302.32 ± 395.23 1201.21 ± 311.91 t=1.171 0.245 IgA (mg/dL) 7.900 (51.00–97.00) 9.210 (59.25–12.82) Z=1.276 0.010 QFA (mg/dL) 2.810.01 (900–31.00) 19.50 (15.67–52.83.50) Z=1.859 0.010 QFA (mg/mL) 6.20 (3.50–9.95) 4.30 (3.00–7.97) Z=2.891 0.016 SVE (ng/mL) 1.20 (1.00–2.27) 1.20 (0.80–2.00) Z=0.585 0.559 SVE (ng/mL) 1.20 (1.00–2.74) 1.20 (0.80–2.00) Z=0.585 0.559 L-4 (pg/mL) 1.20 (1.00–2.74) 1.20 (0.80–3.20) Z=1.442 0.114 L-4 (pg/mL) 1.20 (1.00–2.74) <td>TG (mmoL/L)</td> <td>1.16 (0.87–1.60)</td> <td>0.99 (0.75–1.43)</td> <td>Z=2.053</td> <td>0.040</td>	TG (mmoL/L)	1.16 (0.87–1.60)	0.99 (0.75–1.43)	Z=2.053	0.040
LDL (mmoL/L) 207 (177-28) 231 (184-28) Z=1139 0.120 CRP (mg/L) 8.60 (3.20-20.67) 4.77 (2.45-23.50) Z=1.319 0.187 C4 (mg/dL) 27.66 ± 7.95 2.94.7 ± 9.25 t=0.774 0.441 C3 (mg/dL) 121.32 ± 28.13 1192.82 ± 2.97 t=0.323 0.747 IgG (mg/dL) 130.23 ± 28.13 1201.21 ± 31191 t=1.171 0.245 IgM (mg/dL) 79.00 (51.00-97.00) 92.10 (59.25-127.25) Z=1.561 0.118 CEA (ng/mL) 2.81.00 (190.00-310.00) 195.50 (15.67-283.50) Z=1.891 0.0160 Oxfa 2.1 - 1 (ng/mL) 3.80 (2.20-7.62) 3.00 (1.90-6.70) Z=1.89 0.0066 NSE (ng/mL) 1.84 (0.13.35-2648) 15.60 (12.83-20.00) Z=2.997 0.003 SCC (ng/mL) 1.20 (1.00-2.71) 1.20 (0.80-2.30) Z=1.442 0.119 L2 (ng/mL) 1.12 (0.27-1.83) 1.23 (0.36-2.34) Z=1.490 0.137 L2 (ng/mL) 1.87 (0.33-3.20) 2.00 (1.43-3.297 Z=1.442 0.149 L1-10 (pg/mL) 1.87 (0.35-5.2	TC (mmoL/L)	3.93 (3.30–4.76)	4.17 (3.54–4.74)	Z=1.272	0.203
CRP (mg/L) 8.60 (2.20-20.67) 4.77 (2.45-23.50) Z = 1.319 0.187 Fibringen (g/L) 3.67 (2.87-493) 3.58 (2.96-478) Z=0.232 0.817 C4 (mg/dL) 121.32 ± 28.13 1192.8 ± 22.97 t=0.323 0.747 IgG (mg/dL) 1302.32 ± 395.23 1201.21 ± 31.91 t=1.171 0.245 IgA (mg/dL) 281.00 (190.00-310.00) 195.50 (156.75-283.50) Z=1.561 0.018 CAF (ng/mL) 6.20 (350-9.95) 4.30 (3.00-7.97) Z=2.591 0.010 CySE (ng/mL) 18.0 (2.20-7.62) 3.00 (1.90-67.00) Z=1.839 0.066 SSE (ng/mL) 1.20 (1.00-2.27) 1.20 (0.80-2.00) Z=0.585 0.559 IL-2 (pg/mL) 0.70 (0.31-14.7) 1.02 (0.36-2.34) Z=1.400 0.137 IL-4 (pg/mL) 1.12 (0.27-18.3) 1.23 (0.36-2.34) Z=1.402 0.137 IL-4 (pg/mL) 1.87 (1.03-3.20) 2.00 (1.43-3.97) Z=1.202 0.229 IL-4 (pg/mL) 1.67 (0.36-5.22) 2.70 (0.89-5.63) Z=1.442 0.149 IL-4 (pg/mL) 1.87 (1.0	LDL (mmoL/L)	2.07 (1.77–2.83)	2.31 (1.84–2.83)	Z=1.103	0.270
Fibrinogen (g/L) 3.67 (2.87-4.93) 3.58 (2.96-4.78) Z=0.232 0.817 C4 (mg/dL) 2.766 ±7.95 2.947 ± 9.25 t=0.774 0.441 (3 (mg/dL) 121 32 ± 28.13 119.28 ± 22.97 t=0.323 0.747 IgG (mg/dL) 1302 32 ± 395.23 1201.21 ± 311.91 t=-1.171 0.245 IgM (mg/dL) 281.00 (190.00-310.00) 195.50 (156.75-283.50) Z=1.266 0.010 CFA (ng/mL) 6.20 (350-9.95) 4.30 (3.00-7.97) Z=2.991 0.010 Cyf (a g/mL) 1.840 (13.35-64.86) 1.560 (12.83-0.00) Z=0.585 0.559 IL-2 (pg/mL) 1.00 (10.0-2.27) 1.20 (0.80-2.00) Z=0.480 0.379 IL-6 (pg/mL) 1.02 (0.76-24.79) 7.98 (4.33-22.88) Z=0.880 0.379 IL-16 (pg/mL) 1.87 (10.3-3.20) 2.00 (1.43-3.97) Z=1.442 0.151 IL-16 (pg/mL) 1.87 (10.3-3.20) 2.00 (1.43-3.97) Z=1.441 0.149 IL-16 (pg/mL) 1.87 (10.5-3.21) 2.3 (0.10-12.06) Z=0.926 0.373 IL-16 (pg/mL) 1.87 (CRP (mg/L)	8.60 (3.20–20.67)	4.77 (2.45–23.50)	Z=1.319	0.187
C4 (mg/dL) 27.66±7.95 29.47±9.25 t=0.774 0.441 C3 (mg/dL) 121.32±8.13 119.28±2.97 t=0.323 0.747 IgG (mg/dL) 1302.32±395.23 120.121±311.91 t=1.171 0.245 IgM (mg/dL) 29100 (51.00-97.00) 95.50 (15675-283.50) Z=1.561 0.118 CFA (ng/mL) 6.20 (3.50-99.5) 4.30 (3.00-7.97) Z=2.591 0.010 Cyfra 21-1 (ng/mL) 3.80 (2.02-7.62) 3.00 (1.90-67) Z=1.839 0.666 NSE (ng/mL) 1.840 (13.35-26.48) 1.560 (12.83-20.00) Z=0.585 0.559 IL-2 (pg/mL) 0.70 (0.31-1.47) 1.02 (0.04-1.96) Z=1.442 0.137 IL-4 (pg/mL) 1.12 (0.27-1.83) 1.23 (0.36-2.24) Z=1.049 0.137 IL-4 (pg/mL) 1.027 (5.76-24.79) 7.98 (4.33-22.88) Z=0.880 0.379 IL-10 (pg/mL) 1.67 (0.36-5.22) 2.72 (0.89-5.03) Z=1.441 0.149 IL-17 (pg/mL) 1.67 (0.36-5.22) 2.72 (0.89-5.03) Z=1.441 0.149 IL-10 (pg/mL) 1.67 (0.36-5.22)	Fibrinogen (g/L)	3.67 (2.87–4.93)	3.58 (2.96–4.78)	Z=0.232	0.817
C3 (mg/dL) 12132±28.13 119.28±2.97 t=0.323 0.747 IgG (mg/dL) 1302.32±395.23 12012±11.91 t=1.171 0.245 IgM (mg/dL) 281.00 (190.00-97.00) 92.10 (59.25-127.25) Z=1.561 0.118 CEA (ng/mL) 6.20 (3.50-9.95) 4.30 (3.00-7.97) Z=2.591 0.010 CyTa 1-1 (ng/mL) 3.80 (2.20-7.62) 3.00 (190-6.70) Z=1.839 0.066 NSE (ng/mL) 1.20 (1.00-2.27) 1.20 (0.80-2.00) Z=0.58 0.559 IL-2 (pg/mL) 0.70 (0.31-1.47) 1.02 (0.34-1.96) Z=1.442 0.151 IL-6 (pg/mL) 1.12 (0.27-1.83) 1.23 (0.36-2.34) Z=1.490 0.137 IL-6 (pg/mL) 1.02 (7.57-624.79) 7.98 (4.33-22.88) Z=0.880 0.379 IL-10 (pg/mL) 1.87 (1.03-3.20) 2.200 (1.43-3.97) Z=1.443 0.149 IL-17 (pg/mL) 1.15 (0.10-9.66) 2.23 (0.10-12.06) Z=0.926 0.373 T cells colsh (%b) 67.27±12.58 67.82±10.40 t=0.283 0.778 IL-17 (pg/mL) 1.15 (0.10-9.66) 2.23 (0.10-12.06) Z=0.301 0.379 T cells	C4 (mg/dL)	27.66 ± 7.95	29.47±9.25	t=0.774	0.441
IgG (mg/dL) 1302.32 ± 395.23 1201.21 ± 311.91 t=-1.171 0.245 IgM (mg/dL) 7900 (51.00-97.00) 9210 (52.5-127.25) Z=1.26 0.202 IgA (mg/dL) 281.00 (190.00-310.00) 195.50 (156.75-283.50) Z=1.561 0.118 CFA (ng/mL) 6.20 (3.50-9.95) 4.30 (3.00-7.97) Z=2.597 0.003 CG (ng/mL) 1.84 (13.35-26.48) 15.60 (12.83-20.00) Z=0.585 0.559 L2 (ng/mL) 1.20 (1.00-2.27) 1.20 (0.80-2.00) Z=0.488 0.137 L4 (ng/mL) 1.12 (0.27-1.83) 1.23 (0.36-2.34) Z=1.490 0.137 L9 (ng/mL) 1.87 (1.03-3.20) 2.00 (143-3.97) Z=1.420 0.137 L9 (ng/mL) 1.87 (0.36-5.22) 2.72 (0.89-5.03) Z=1.441 0.149 LN-Y (ng/mL) 1.67 (0.36-5.22) 2.72 (0.89-5.03) Z=1.433 0.149 LN-Y (ng/mL) 1.67 (0.36-5.22) 2.72 (0.89-5.03) Z=1.431 0.149 LN-Y (ng/mL) 1.67 (0.36-5.22) 2.72 (0.89-5.03) Z=1.431 0.149 LN-Y (ng/mL) 1.67 (0	C3 (mg/dL)	121.32±28.13	119.28±22.97	t=-0.323	0.747
IgM (mg/dL) 79.00 (51.00–97.00) 92.10 (59.25–127.25) Z = 1.276 0.020 IgA (mg/dL) 281.00 (190.00–310.00) 195.00 (156.75–283.50) Z = 1.561 0.118 CEA (ng/mL) 6.20 (350–9.95) 3.00 (1.90–6.70) Z = 2.591 0.000 Oxfa 21 – 1 (ng/mL) 3.80 (2.20–7.62) 3.00 (1.90–6.70) Z = 2.897 0.003 SEC (ng/mL) 1.84 (01335–26.48) 15.60 (12.83–20.00) Z = 2.997 0.003 SCC (ng/mL) 1.20 (1.00–2.27) 1.20 (0.80–2.00) Z = 1.42 0.151 L4 (pg/mL) 1.12 (0.27–1.83) 1.23 (0.36–2.34) Z = 1.402 0.151 L4 (pg/mL) 1.027 (5.76–24.79) 7.98 (4.33–22.88) Z = 0.880 0.379 L1-10 (pg/mL) 1.87 (1.03–3.20) 2.00 (1.43–3.97) Z = 1.420 0.129 NF-q (pg/mL) 1.15 (0.10–9.66) 2.23 (0.10–12.06) Z = 0.333 T = 1.420 0.373 T cells count (CD3+/cell/µL) 917.50 (715.50–1105.75) 854.00 (571.75–112.50) Z = 1.431 0.149 L17 A (pg/mL) 1.50 (10–9.66) 2.23 (0.10–12.06) Z = 0.3	lgG (mg/dL)	1302.32±395.23	1201.21±311.91	t=-1.171	0.245
IgA (mg/dL) 281.00 (190.00-310.00) 195.50 (156.75-283.50) Z = 1.561 0.118 CFA (ng/mL) 6.20 (350-9.95) 4.30 (3.00-7.97) Z = 2.591 0.010 Cyfra 21 - 1 (ng/mL) 3.80 (2.20-7.62) 3.00 (1.90-6.77) Z = 1.839 0.066 SSE (ng/mL) 1.840 (1335-26.48) 15.60 (12.83-20.00) Z = 2.997 0.003 SCC (ng/mL) 1.20 (1.00-2.27) 1.20 (0.80-2.00) Z = 1.442 0.151 Li-4 (pg/mL) 1.12 (0.27-1.83) 1.23 (0.36-2.34) Z = 1.490 0.137 Li-6 (pg/mL) 1.027 (5.76-24.79) 7.98 (4.33-2.288) Z = 0.880 0.379 IL-10 (pg/mL) 1.027 (0.36-2.21) 2.00 (1.43-3.97) Z = 1.420 0.137 IL-10 (pg/mL) 1.67 (0.36-5.22) 2.72 (0.89-5.03) Z = 1.443 0.149 IL-17 (pg/mL) 1.15 (0.10-9.66) 2.23 (0.10-12.06) Z = 0.926 0.373 T cells (CD3+) (cell/µL) 91.75 0 (715.50-1105.75) 854.00 (571.75-112.50) Z = 1.350 0.176 ILe17 cells count (CD3+,CD4+) (cell/µL) 52.000 (34.800-665.50) 477.50 (32.05-631.25)	lgM (mg/dL)	79.00 (51.00–97.00)	92.10 (59.25–127.25)	Z=1.276	0.202
CEA (ng/mL) 6.20 (3.50-9.95) 4.30 (3.00-7.97) Z=2.591 0.010 Cyfra J - 1 (ng/mL) 3.80 (2.20-7.62) 3.00 (1.90-6.70) Z=1.839 0.066 NSE (ng/mL) 1.840 (13.35-2648) 15.60 (12.83-2000) Z=2.997 0.003 SCC (ng/mL) 1.20 (1.00-2.27) 1.20 (0.80-2.00) Z=1.442 0.151 L4 (pg/mL) 1.12 (0.27-1.83) 1.23 (0.36-2.34) Z=1.490 0.337 L1-6 (pg/mL) 1.02 (7.57-24.79) 7.98 (4.33-22.88) Z=0.880 0.379 L1-10 (pg/mL) 1.87 (10.3-3.20) 2.00 (1.43-3.97) Z=1.443 0.149 INF-a (pg/mL) 1.67 (0.36-5.22) 2.72 (0.89-5.03) Z=1.441 0.149 IL-17 A (pg/mL) 1.15 (0.10-9.66) 2.23 (0.10-12.06) Z=0.926 0.373 T cells (CD3+) (%) 67.27 ± 12.58 67.82 ± 10.40 t=0.283 0.778 T cells (CD3+) (%) 67.27 ± 12.58 67.82 ± 10.40 t=0.283 0.778 C cells count (CD3+,CD4+)(cell/µL) 52.00 (34.80-665.50) 477.50 (32.50-613.25) Z=0.880 0.379 <t< td=""><td>lgA (mg/dL)</td><td>281.00 (190.00-310.00)</td><td>195.50 (156.75–283.50)</td><td>Z=1.561</td><td>0.118</td></t<>	lgA (mg/dL)	281.00 (190.00-310.00)	195.50 (156.75–283.50)	Z=1.561	0.118
Cyfa 21 - 1 (ng/mL) 3.80 (2.20 - 7.62) 3.00 (1.90 - 6.70) Z = 1.839 0.066 NSE (ng/mL) 18.40 (13.35 - 26.48) 15.60 (12.83 - 20.00) Z = 2.997 0.003 SCC (ng/mL) 1.20 (1.00 - 2.27) 1.20 (0.80 - 2.00) Z = 1.442 0.151 L4 (ng/mL) 1.12 (0.27 - 1.83) 1.23 (0.36 - 2.34) Z = 1.409 0.137 L1-6 (ng/mL) 10.27 (5.76 - 24.79) 7.98 (4.33 - 22.88) Z = 0.880 0.379 L1-10 (ng/mL) 1.87 (1.03 - 3.20) 2.00 (1.43 - 3.97) Z = 1.443 0.149 FN-γ (ng/mL) 1.67 (0.36 - 5.22) 2.72 (0.89 - 5.03) Z = 1.441 0.149 L1-7 A (ng/mL) 1.15 (0.10 - 9.66) 2.23 (0.10 - 12.06) Z = 0.323 0.778 T cells (CD3+() (cell/µL) 917.50 (715.50 - 1105.75) 854.00 (571.75 - 112.50) Z = 1.350 0.177 Helper T cells 36.41 ± 11.82 39.16 ± 10.53 t = 1.420 0.158 (CD3+, CD4+) (cell/µL) 52.000 (34.800 - 665.50) 477.50 (32.050 - 631.25) Z = 0.830 0.377 B cells Count (CD3+, CD4+) (cell/µL) 52.000 (34.800 - 665.50)	CEA (ng/mL)	6.20 (3.50–9.95)	4.30 (3.00–7.97)	Z=2.591	0.010
NSE (ng/mL) 18.40 (13.35–26.48) 15.60 (12.83–20.00) Z=2.997 0.003 SCC (ng/mL) 1.20 (1.00–2.27) 1.20 (0.80–2.00) Z=0.585 0.559 IL-2 (pg/mL) 0.70 (0.31–1.47) 1.02 (0.34–1.96) Z=1.490 0.137 IL-6 (pg/mL) 1.12 (0.27–1.83) 1.23 (0.36–2.34) Z=0.880 0.379 IL-10 (pg/mL) 1.87 (1.03–3.20) 2.00 (1.43–3.97) Z=1.420 0.129 TNF-0 (pg/mL) 1.67 (0.36–5.22) 2.272 (0.89–5.03) Z=1.441 0.149 IL-17 A (pg/mL) 1.15 (0.10–9.66) 2.23 (0.10–12.06) Z=0.320 0.778 T cells (CD3+) (%) 67.27±12.58 67.82±10.40 t=0.283 0.778 T cells (CD3+) (cell/µL) 917.50 (715.50–1105.75) 854.00 (571.75–1122.50) Z=1.350 0.177 Helper T cells 3.641±11.82 39.16±1.053 t=1.420 0.158 (CD3+, CD4+) (cell/µL) 52.00.01348.00–665.50 Z=0.313 0.897 B cells Count (CD3+, CD4+) (cell/µL) 52.00.01348.00–665.50 Z=0.313 0.897 CD19+) (%) 8.78 (5.09–13.	Cyfra 21 – 1 (ng/mL)	3.80 (2.20–7.62)	3.00 (1.90–6.70)	Z=1.839	0.066
SCC (ng/mL) 1.20 (1.00–2.27) 1.20 (0.80–2.00) Z=0585 0.599 IL-2 (ng/mL) 0.70 (0.31–1.47) 1.02 (0.34–1.96) Z=1.442 0.151 IL-4 (ng/mL) 1.12 (0.27–1.83) 1.23 (0.36–2.34) Z=1.080 0.379 IL-6 (ng/mL) 1.07 (5.76–24.79) 7.98 (4.33–22.88) Z=0.080 0.379 IL-10 (ng/mL) 1.87 (1.03–3.20) 2.00 (1.43–3.97) Z=1.022 0.229 INF-a (ng/mL) 1.67 (0.36–5.22) 2.72 (0.89–5.03) Z=1.443 0.149 IL-17 A (ng/mL) 1.15 (0.10–9.66) 2.23 (0.10–12.06) Z=0.926 0.373 T cells cO1+()(%h) 67.27 ± 1.258 67.82 ± 10.40 t=0.283 0.778 T cells count (CD3+,()cell/µL) 91.750 (71.5.0 – 110.57) 854.00 (571.7.5 – 112.50) Z=1.350 0.177 Helper T cells count (CD3+,CD4+)(cell/µL) 92.000 (348.00–665.50) 477.50 (32.0.5.0–631.25) Z=0.880 0.379 B cells count (CD3+,CD4+)(cell/µL) 52.000 (348.00–665.50) 477.50 (32.0.5.0–631.25) Z=0.880 0.371 Killer T cells count (CD3+,CD4+)(cell/µL) 52.000 (348.00–665.50)	NSE (ng/mL)	18.40 (13.35–26.48)	15.60 (12.83–20.00)	Z=2.997	0.003
IL-2 (pg/mL) 0.70 (0.31–1.47) 1.02 (0.34–1.96) Z=1.442 0.151 IL-4 (pg/mL) 1.12 (0.27–1.83) 1.23 (0.36–2.34) Z=1.490 0.137 IL-6 (pg/mL) 10.27 (5.76–24.79) 7.98 (4.33–22.88) Z=0.880 0.379 IL-10 (pg/mL) 1.87 (1.03–3.20) 2.00 (1.43–3.97) Z=1.202 0.229 TNF-a (pg/mL) 1.67 (0.36–5.22) 2.49 (1.17–4.89) Z=1.443 0.149 IL-17 A (pg/mL) 1.15 (0.10–9.66) 2.23 (0.10–12.06) Z=0.926 0.379 T cells (CD3+) (%) 67.27 ± 12.58 67.82 ± 10.40 t=0.283 0.778 T cells count (CD3+)(cell/µL) 917.50 (715.50–1105.75) 854.00 (571.75–1122.50) Z=1.350 0.177 Helper T cells count (CD3+,CD4+)(cell/µL) 520.00 (348.00–665.50) 477.50 (320.50–631.25) Z=0.880 0.379 B cells (CD1+) (%) 8.78 (5.09–13.40) 8.48 (5.50–12.22) Z=0.130 0.897 B cells (CD1+,(D5+,CD4+)(cell/µL) 367.50 (230.50–490.75) 281.50 (186.75–415.75) Z=2.193 0.028 Killer T cells (cont (CD3+,CD8+) (cell/µL) 367.50 (230.50–490.75) <td>SCC (ng/mL)</td> <td>1.20 (1.00–2.27)</td> <td>1.20 (0.80–2.00)</td> <td>Z=0.585</td> <td>0.559</td>	SCC (ng/mL)	1.20 (1.00–2.27)	1.20 (0.80–2.00)	Z=0.585	0.559
IL-4 (pg/mL) 1.12 (0.27–1.83) 1.23 (0.36–2.34) Z=1.490 0.137 IL-6 (pg/mL) 10.27 (5.76–24.79) 7.98 (4.33–22.88) Z=0.880 0.379 IL-10 (pg/mL) 1.87 (1.03–3.20) 2.00 (1.43–3.97) Z=1.202 0.229 TNF-a (pg/mL) 2.04 (0.99–3.72) 2.49 (1.17–4.89) Z=1.443 0.149 IFN-y (pg/mL) 1.67 (0.36–5.22) 2.72 (0.89–5.03) Z=1.441 0.149 IL-17 A (pg/mL) 1.15 (0.10–9.66) 2.23 (0.10–12.06) Z=0.926 0.378 T cells (CD3+) (%) 67.27 ± 12.58 67.82 ± 10.40 t=0.283 0.778 T cells count (CD3+)(cell/µL) 917.50 (715.50–1105.75) 854.00 (571.75–1122.50) Z=1.350 0.177 Helper T cells count (CD3+,CD4+)(cell/µL) 520.00 (348.00–665.50) 477.50 (320.50–631.25) Z=0.880 0.379 B cells (CD1+y) (%) 8.78 (5.09–13.40) 8.48 (5.0–12.22) Z=0.130 0.897 B cells count (CD3+,CD4+)(cell/µL) 520.00 (348.00–665.50) 477.50 (320.50–631.25) Z=0.884 0.379 B cells count (CD3+,CD4+)(cell/µL) 520.00 (348.00–665.50) 477.50 (320.50–631.25) Z=0.9130 0.897	IL-2 (pg/mL)	0.70 (0.31–1.47)	1.02 (0.34–1.96)	Z=1.442	0.151
LL-6 (pg/mL) 10.27 (576-24.79) 7.98 (4.33-22.88) Z=0.880 0.379 LL-10 (pg/mL) 1.87 (1.03-3.20) 2.00 (1.43-3.97) Z=1.202 0.229 TNF-a (pg/mL) 2.04 (0.99-3.72) 2.49 (1.17-4.89) Z=1.443 0.149 IFN-y (pg/mL) 1.67 (0.36-5.22) 2.72 (0.89-5.03) Z=1.441 0.149 IL-17 A (pg/mL) 1.15 (0.10-9.66) 2.23 (0.10-12.06) Z=0.926 0.373 T cells (CD3+) (%) 67.27 ± 12.58 67.82 ± 10.40 t=0.283 0.778 T cells count (CD3+)(cell/µL) 917.50 (715.50-1105.75) 854.00 (571.75-1122.50) Z=1.350 0.177 Heiper T cells count (CD3+,CD4+)(cell/µL) 917.50 (715.50-1105.75) 854.00 (571.75-1122.50) Z=0.880 0.379 B cells count (CD3+,CD4+)(cell/µL) 520.00 (348.00-665.50) 477.50 (320.50-631.25) Z=0.880 0.377 B cells (CD19+) (%) 8.78 (5.09-13.40) 8.48 (5.50-12.22) Z=0.130 0.897 B cells (CD19+, (MD4+)(CD3+,CD4+)(cell/µL) 367.50 (230.50-430.75) 24.10 (18.24-32.07) Z=0.971 0.331 Killer T cells count (CD3+,CD8+) (sell/µL)<	IL-4 (pg/mL)	1.12 (0.27–1.83)	1.23 (0.36–2.34)	Z=1.490	0.137
IL-10 (pg/mL) 1.87 (1.03-3.20) 2.00 (1.43-3.97) Z=1.202 0.292 TNF-a (pg/mL) 2.04 (0.99-3.72) 2.49 (1.17-4.89) Z=1.443 0.149 IFN-y (pg/mL) 1.67 (0.36-5.22) 2.72 (0.89-5.03) Z=1.441 0.149 IL-17 A (pg/mL) 1.15 (0.10-9.66) 2.23 (0.10-12.06) Z=0.926 0.373 T cells (CD3+) (%) 67.27 ± 12.58 67.82 ± 10.40 t=0.283 0.778 T cells count (CD3+)(cell/µL) 917.50 (715.50-1105.75) 854.00 (571.75-1122.50) Z=1.350 0.177 Helper T cells count (CD3+,CD4+)(cell/µL) 917.50 (715.50-1105.75) 854.00 (571.75-1122.50) Z=0.880 0.379 B cells count (CD3+,CD4+)(cell/µL) 92.000 (348.00-665.50) 477.50 (320.50-631.25) Z=0.880 0.379 B cells count (CD3+,CD4+)(cell/µL) 52.000 (348.00-665.50) 477.50 (320.50-631.25) Z=0.130 0.897 B cells (CD19+) (%) 8.78 (5.09-13.40) 8.48 (5.50-12.22) Z=0.130 0.897 B cells count (CD3+,CD8+) (cell/µL) 367.50 (230.50-490.75) 281.50 (186.75-415.75) Z=2.193 0.028 Killer T cells	IL-6 (pg/mL)	10.27 (5.76–24.79)	7.98 (4.33–22.88)	Z=0.880	0.379
TNF-a (pg/mL) 2.04 (0.99-3.72) 2.49 (1.17-4.89) Z = 1.43 0.149 IFN-y (pg/mL) 1.67 (0.36-5.22) 2.72 (0.89-5.03) Z = 1.441 0.149 IL-17 A (pg/mL) 1.15 (0.10-9.66) 2.23 (0.10-12.06) Z = 0.926 0.373 T cells (CD3+) (%) 67.27 ± 12.58 67.82 ± 10.40 t= 0.283 0.778 T cells count (CD3+)(cell/µL) 917.50 (715.50-1105.75) 854.00 (571.75-1122.50) Z = 1.350 0.177 Helper T cells 36.41 ± 11.82 39.16 ± 10.53 t= 1.420 0.158 (CD3+,CD4+) (%) 87.8 (5.09-13.40) 84.8 (5.50-12.22) Z = 0.130 0.897 B cells count (CD3+,CD4+)(cell/µL) 52.000 (34.8.00-665.50) 477.50 (320.50-631.25) Z = 0.880 0.379 B cells count (CD3+,CD4+)(cell/µL) 52.000 (34.8.00-665.50) 477.50 (320.50-631.25) Z = 0.130 0.897 B cells (CD19+) (%) 8.78 (5.09-13.40) 8.48 (5.50-12.22) Z = 0.130 0.897 B cells count (CD3+,CD8+) (%) 2.862 (17.56-34.87) 24.10 (18.24-32.07) Z = 0.971 0.331 Killer T cells count (CD3+,CD8+	IL-10 (pg/mL)	1.87 (1.03–3.20)	2.00 (1.43–3.97)	Z=1.202	0.229
IFN-y (pg/mL) 1.67 (0.36–5.22) 2.72 (0.89–5.03) Z = 1.41 0.149 IL-17 A (pg/mL) 1.15 (0.10–9.66) 2.23 (0.10–12.06) Z = 0.926 0.373 T cells (CD3+) (%) 67.27 ± 12.58 67.82 ± 10.40 t = 0.283 0.778 T cells count (CD3+)(cell/µL) 917.50 (715.50–1105.75) 854.00 (571.75–1122.50) Z = 1.350 0.177 Helper T cells 36.41 ± 11.82 39.16 ± 10.53 t = 1.420 0.158 (CD3+,CD4+) (%) 8.78 (5.09–13.40) 8.48 (5.50–12.22) Z = 0.880 0.379 B cells (CD19+) (%) 8.78 (5.09–13.40) 8.48 (5.50–12.22) Z = 0.84 0.377 (CD19+) (%) 8.78 (5.09–13.40) 8.48 (5.50–12.22) Z = 0.84 0.377 B cells count 110.00 (64.00–192.75) 96.00 (60.75–148.50) Z = 0.84 0.377 (CD19+) (%) 2.862 (17.56–34.87) 2.410 (18.24–32.07) Z = 0.971 0.318 Killer T cells count (CD3+,CD8+) (cell/µL) 367.50 (230.50–490.75) 281.50 (186.75–415.75) Z = 2.193 0.028 NK cells count (CD3+,CD8+) (cell/µL) 367.50 (230.50	TNF-α (pg/mL)	2.04 (0.99–3.72)	2.49 (1.17–4.89)	Z=1.443	0.149
IL-17 A (pg/mL)1.15 (0.10–9.66)2.23 (0.10–12.06)Z = 0.9260.373T cells (CD3+) (%)67.27 ± 12.5867.82 ± 10.40t = 0.2830.778T cells count (CD3+)(cell/µL)917.50 (715.50–1105.75)854.00 (571.75–1122.50)Z = 1.3500.177Helper T cells36.41 ± 11.8239.16 ± 10.53t = 1.4200.158(CD3+,CD4+) (%)520.00 (348.00–665.50)477.50 (320.50–631.25)Z = 0.8800.379B cells (CD19+) (%)8.78 (5.09–13.40)8.48 (5.50–12.22)Z = 0.1300.897B cells (CD19+) (%)8.78 (5.09–13.40)8.48 (5.50–12.22)Z = 0.9710.331Killer T cells (CD3+,CD8+) (%)28.62 (17.56–34.87)24.10 (18.24–32.07)Z = 0.9710.331Killer T cells count (CD3+,CD8+) (cell/µL)367.50 (230.50–490.75)281.50 (186.75–415.75)Z = 2.1930.028NK cells count (CD3+,CD8+) (cell/µL)367.50 (230.50–490.75)281.50 (186.75–415.75)Z = 0.9710.331Killer T cells count (CD3+,CD8+) (cell/µL)367.50 (230.50–490.75)281.50 (186.75–415.75)Z = 2.1930.028NK cells count (CD3+,CD5+) (cell/µL)19.45 (12.85–30.20)20.00 (13.98–29.88)Z = 0.3530.724NK cells count (CD3+,CD5+) (cell/µL)1382.50 (1189.25–1731.25)1232.00 (894.75–1635.00)Z = 1.8670.062(CD4+) (cell/µL)1.65 (1.03–2.34)Z = 1.1350.256	IFN-γ (pg/mL)	1.67 (0.36–5.22)	2.72 (0.89–5.03)	Z=1.441	0.149
T cells (CD3+) (%) 67.27 ± 12.58 67.82 ± 10.40 t = 0.283 0.778 T cells count (CD3+)(cell/µL) 917.50 (715.50-1105.75) 854.00 (571.75-1122.50) Z = 1.350 0.177 Helper T cells 36.41 ± 11.82 39.16 ± 10.53 t = 1.420 0.158 (CD3+,CD4+) (%) 77.50 (320.50-631.25) Z = 0.880 0.379 B cells (CD19+) (%) 8.78 (5.09-13.40) 8.48 (5.50-12.22) Z = 0.130 0.897 B cells count 110.00 (64.00-192.75) 96.00 (60.75-148.50) Z = 0.971 0.331 Killer T cells count (CD3+,CD8+) (%) 28.62 (17.56-34.87) 24.10 (18.24-32.07) Z = 0.971 0.331 Killer T cells count (CD3+,CD8+) (%) 19.45 (12.85-30.20) 20.00 (13.98-29.88) Z = 0.353 0.724 NK cells count 256.50 (153.50-413.25) 241.50 (147.50-343.50) Z = 0.917 0.359 (CD16+, CD56+) (cell/µL) 1082.50 (1189.25-1731.25) 1232.00 (894.75-1635.00) Z = 1.867 0.062 (CD4+/ CD8+ratio 1.44 (0.95-2.14) 1.65 (1.03-2.34) Z = 1.135 0.256	IL-17 A (pg/mL)	1.15 (0.10–9.66)	2.23 (0.10-12.06)	Z=0.926	0.373
T cells count (CD3+)(cell/µL) 917.50 (715.50–1105.75) 854.00 (571.75–1122.50) Z=1.350 0.177 Helper T cells (CD3+,CD4+) (%) 36.41±11.82 39.16±10.53 t=1.420 0.158 Helper T cells count (CD3+,CD4+)(cell/µL) 520.00 (348.00–665.50) 477.50 (320.50–631.25) Z=0.880 0.379 B cells (CD19+) (%) 8.78 (5.09–13.40) 8.48 (5.50–12.22) Z=0.130 0.897 B cells count 110.00 (64.00–192.75) 96.00 (60.75–148.50) Z=0.884 0.377 (CD19+)(cell/µL) 28.62 (17.56–34.87) 24.10 (18.24–32.07) Z=0.971 0.331 Killer T cells count (CD3+,CD8+) (cell/µL) 367.50 (230.50–490.75) 281.50 (186.75–415.75) Z=2.193 0.028 NK cells count (CD3+,CD8+) (cell/µL) 367.50 (230.50–490.75) 241.00 (18.24–32.07) Z=0.971 0.331 Killer T cells (cD16+, CD56+) (%) 19.45 (12.85–30.20) 20.00 (13.98–29.88) Z=0.353 0.724 NK cells count 256.50 (153.50–413.25) 241.50 (147.50–343.50) Z=0.917 0.359 (CD16+, CD56+) (cell/µL) 1582.50 (1189.25–1731.25) 1232.00 (894.75–1635.00) Z=1.867 0.062 (CD4+/ CD8+ ratio 1.44 (095–2.14)	T cells (CD3+) (%)	67.27 ± 12.58	67.82±10.40	t=0.283	0.778
Helper T cells36.41 ± 11.8239.16 ± 10.53t = 1.4200.158(CD3+,CD4+) (%)520.00 (348.00–665.50)477.50 (320.50–631.25)Z = 0.8800.379B cells (CD19+) (%)8.78 (5.09–13.40)8.48 (5.50–12.22)Z = 0.1300.897B cells count110.00 (64.00–192.75)96.00 (60.75–148.50)Z = 0.8840.377(CD19+) (cell/µL)28.62 (17.56–34.87)24.10 (18.24–32.07)Z = 0.9710.331Killer T cells count (CD3+,CD8+) (cell/µL)367.50 (230.50–490.75)281.50 (186.75–415.75)Z = 2.1930.028NK cells count (CD3+,CD8+) (cell/µL)367.50 (230.50–490.75)20.00 (13.98–29.88)Z = 0.3530.724NK cells count (CD3+,CD8+) (cell/µL)19.45 (12.85–30.20)20.00 (13.98–29.88)Z = 0.9170.359(CD16+, CD56+) (cell/µL)1382.50 (1189.25–1731.25)241.50 (147.50–343.50)Z = 0.9170.359(CD16+, CD56+) (cell/µL)1382.50 (1189.25–1731.25)1232.00 (894.75–1635.00)Z = 1.8670.062(CD4+/ CD8+ ratio1.44 (0.95–2.14)1.65 (1.03–2.34)Z = 1.1350.256	T cells count (CD3+)(cell/µL)	917.50 (715.50–1105.75)	854.00 (571.75–1122.50)	Z=1.350	0.177
(CD3+,CD4+) (%)Helper T cells count (CD3+,CD4+)(cell/µL)520.00 (348.00-665.50)477.50 (320.50-631.25)Z = 0.8800.379B cells (CD19+) (%)8.78 (5.09-13.40)8.48 (5.50-12.22)Z = 0.1300.897B cells count110.00 (64.00-192.75)96.00 (60.75-148.50)Z = 0.8840.377(CD19+)(cell/µL)28.62 (17.56-34.87)24.10 (18.24-32.07)Z = 0.9710.331Killer T cells (CD3+,CD8+) (%)28.62 (17.56-34.87)24.10 (18.24-32.07)Z = 0.9710.331Killer T cells count (CD3+,CD8+) (cell/µL)367.50 (230.50-490.75)281.50 (186.75-415.75)Z = 2.1930.028NK cells (CD16+, CD56+) (%)19.45 (12.85-30.20)20.00 (13.98-29.88)Z = 0.3530.724NK cells count256.50 (153.50-413.25)241.50 (147.50-343.50)Z = 0.9170.359(CD16+, CD56+) (cell/µL)1382.50 (1189.25-1731.25)1232.00 (894.75-1635.00)Z = 1.8670.062(CD4+/ CD8+ ratio1.44 (0.95-2.14)1.65 (1.03-2.34)Z = 1.1350.256	Helper T cells	36.41±11.82	39.16±10.53	t=1.420	0.158
Helper T cells count (CD3+,CD4+)(cell/µL)520.00 (348.00-665.50)477.50 (320.50-631.25)Z = 0.8800.379B cells (CD19+) (%)8.78 (5.09-13.40)8.48 (5.50-12.22)Z = 0.1300.897B cells count110.00 (64.00-192.75)96.00 (60.75-148.50)Z = 0.8840.377(CD19+)(cell/µL)28.62 (17.56-34.87)24.10 (18.24-32.07)Z = 0.9710.331Killer T cells (CD3+,CD8+) (%)28.62 (17.56-34.87)24.10 (18.24-32.07)Z = 0.9710.331Killer T cells count (CD3+,CD8+) (cell/µL)367.50 (230.50-490.75)281.50 (186.75-415.75)Z = 2.1930.028NK cells (CD16+, CD56+) (%)19.45 (12.85-30.20)20.00 (13.98-29.88)Z = 0.3530.724NK cells count256.50 (153.50-413.25)241.50 (147.50-343.50)Z = 0.9170.359(CD16+, CD56+) (cell/µL)1382.50 (1189.25-1731.25)1232.00 (894.75-1635.00)Z = 1.8670.062(CD4+/ CD8+ ratio1.44 (0.95-2.14)1.65 (1.03-2.34)Z = 1.1350.256	(CD3+,CD4+) (%)				
B cells (CD19+) (%) 8.78 (5.09-13.40) 8.48 (5.50-12.22) Z = 0.130 0.897 B cells count 110.00 (64.00-192.75) 96.00 (60.75-148.50) Z = 0.884 0.377 (CD19+)(cell/µL)	Helper T cells count (CD3+,CD4+)(cell/µL)	520.00 (348.00–665.50)	477.50 (320.50–631.25)	Z=0.880	0.379
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Killer T cells count (CD3+,CD8+) (cell/µL) 367.50 (230.50-490.75) 281.50 (186.75-415.75) Z = 2.193 0.028 NK cells (CD16+, CD56+) (%) 19.45 (12.85-30.20) 20.00 (13.98-29.88) Z = 0.353 0.724 NK cells count 256.50 (153.50-413.25) 241.50 (147.50-343.50) Z = 0.917 0.359 (CD16+, CD56+) (cell/µL) 1382.50 (1189.25-1731.25) 1232.00 (894.75-1635.00) Z = 1.867 0.062 (CD45+)(cell/µL) 1.44 (0.95-2.14) 1.65 (1.03-2.34) Z = 1.135 0.256	Killer T cells (CD3+,CD8+) (%)	28.62 (17.56–34.87)	24.10 (18.24-32.07)	Z=0.971	0.331
NK cells (CD16+, CD56+) (%) 19.45 (12.85-30.20) 20.00 (13.98-29.88) Z=0.353 0.724 NK cells count 256.50 (153.50-413.25) 241.50 (147.50-343.50) Z=0.917 0.359 (CD16+, CD56+) (cell/µL) 1382.50 (1189.25-1731.25) 1232.00 (894.75-1635.00) Z=1.867 0.062 (CD45+)(cell/µL) 1.44 (0.95-2.14) 1.65 (1.03-2.34) Z=1.135 0.256	Killer T cells count (CD3+,CD8+) (cell/µL)	367.50 (230.50-490.75)	281.50 (186.75-415.75)	Z=2.193	0.028
NK cells count 256.50 (153.50-413.25) 241.50 (147.50-343.50) Z = 0.917 0.359 (CD16+, CD56+) (cell/µL) 1382.50 (1189.25-1731.25) 1232.00 (894.75-1635.00) Z = 1.867 0.062 (CD45+)(cell/µL) 1.44 (0.95-2.14) 1.65 (1.03-2.34) Z = 1.135 0.256	NK cells (CD16+, CD56+) (%)	19.45 (12.85–30.20)	20.00 (13.98–29.88)	Z=0.353	0.724
Lymphocyte count 1382.50 (1189.25–1731.25) 1232.00 (894.75–1635.00) Z=1.867 0.062 (CD45+)(cell/µL) 1.65 (1.03–2.34) Z=1.135 0.256	NK cells count	256.50 (153.50–413.25)	241.50 (147.50–343.50)	Z=0.917	0.359
CD4+/CD8+ratio 1.44 (0.95-2.14) 1.65 (1.03-2.34) Z=1.135 0.256	Lymphocyte count	1382.50 (1189.25–1731.25)	1232.00 (894.75–1635.00)	Z=1.867	0.062
	CD4+/CD8+ratio	1.44 (0.95–2.14)	1.65 (1.03–2.34)	Z=1.135	0.256

WBC: white blood cells; PLT: platelets; ANC: absolute neutrophil count; ALC: absolute lymphocyte count; TG: triglycerides; TC: total cholesterol; LDL: low-density lipoprotein; CRP: C-reactive protein; C4: complement 4; C3: complement 3; IgG: immunoglobulin G; IgM: immunoglobulin M; IgA: immunoglobulin A; CEA: carcinoembryonic antigen; Cyfra 21–1: cytokeratin 19 fragment; NSE: neuron-specific enolase; SCC: squamous cell carcinoma antigen; IL-2: interleukin 2; IL-4: interleukin 4; IL-6: interleukin 6; IL-10: interleukin 10; TNF-a: tumor necrosis factor alpha; IFN-y: interferon gamma; IL-17 A: interleukin 17 A

AEs. Statistical analysis indicated no significant difference between the two groups in terms of AEs (p=0.248, $\chi^2=1.337$).

The CPFE+LC group exhibited a higher incidence of interstitial pneumonia (50.0%) and pituitary insufficiency (20.0%). Conversely, the COPD+LC group predominantly experienced interstitial pneumonia (36.8%) and liver dysfunction (15.8%). The incidence and specific

immune-related adverse events in the two groups are presented in Table 4.

Discussion

This retrospective study reveals the following findings: The CPFE+LC group has higher levels of BMI, TG, CEA, NSE, and Killer T cells count compared to the COPD+LC group. Lung cancer in the CPFE group primarily manifests as adenocarcinoma and squamous cell

Kaplan-Meier survival estimates



Fig. 1 Comparison of PFS between lung cancer patients with CPFE or $\ensuremath{\mathsf{COPD}}$



Kaplan-Meier survival estimates

Fig. 2 Comparison of PFS between NSCLC patients with CPFE or COPD



Kaplan-Meier survival estimates

Fig. 3 Comparison of PFS between SCLC patients with CPFE or COPD

carcinoma, predominantly located in the lower lung lobes, with a higher rate of tumor metastasis and a main characteristic of paraseptal emphysema. In contrast, lung cancer in the COPD group is mainly squamous cell carcinoma, mainly found in the upper lung lobes, and associated with centrilobular emphysema. In advanced cancer patients with TNM staging ranging from IIIB to IV, the CPFE+LC group has a shorter median PFS than the COPD+LC group after first-line treatment. Patients with either NSCLC or SCLC who have combined CPFE have a worse prognosis than those with combined COPD. Within the CPFE+LC group, patients may not derive additional benefits from immunotherapy. And there is no significant difference in the incidence of immune-related AEs between the CPFE+LC group and the COPD+LC group receiving immunotherapy.

After conducting a comparative analysis of CPFE+LC and COPD+LC patients, we noticed that the primary common characteristics in both groups were elderly male patients, with the majority having a history of smoking. Additionally, we observed relatively higher BMI and TG levels in the CPFE+LC group. This finding sparked our interest as it may be related to the presence of pulmonary fibrosis. Seeliger et al.'s study [15] also mentioned significantly elevated triglyceride levels in patients associated with interstitial lung diseases, such as pulmonary fibrosis. This provides some support for our observations. Furthermore, the treatment of pulmonary fibrosis may involve the use of glucocorticoids, which could also be linked to the higher BMI and TG levels.

Smoking can impact lung function and affect the clinical symptoms of patients with CPFE and COPD [16, 17]. During our investigation into the differences in lung function between CPFE+LC and COPD+LC patients, we observed that CPFE+LC group had lower values for indicators such as DLCO, DLCO/ VA, RV, and RV/ TLC compared to the COPD+LC group. This indicated a predominant impairment in gas exchange, which is typically associated with pulmonary fibrosis and emphysema. Conversely, the COPD+LC group exhibited worse results in parameters such as FEV1 and FEV1/ FVC. These lung function results reflect the characteristics of the two primary diseases, suggesting that the presence of lung cancer does not lead to significant changes in lung function.

The increased susceptibility of lung cancer in CPFE or COPD patients may primarily be attributed to a combination of factors such as chronic inflammation [18–20], DNA damage [21], and impaired apoptosis function [22, 23]. Additionally, genetic predisposition and occupational exposures [24] can also influence the development of lung cancer. Our research findings suggested that the majority of lung cancer patients with CPFE or COPD have a history of smoking. Smoking-induced oxidative stress can lead to lipid peroxidation and DNA damage

Kaplan-Meier survival estimates



Fig. 4 Comparative analysis of PFS with different treatment modalities in CPFE+LC patients

[25]. It can also induce global changes in gene methylation status and potentially impact genes involved in cell cycle regulation, airway remodeling, wound healing, and more [26, 27], thus promoting carcinogenesis and increasing the risk of lung cancer in patients with a smoking history.

Our study indicated that the proportions of adenocarcinoma (36.00%) and squamous cell carcinoma (34.67%) were quite similar among CPFE patients, while in COPD patients, squamous cell carcinoma was the predominant lung cancer type, accounting for 52.75% of cases. Consistent with our findings, a study by Usui et al. [28] in Japan on CPFE patients with concurrent lung cancer also found a higher proportion of adenocarcinoma (45.5%) compared to squamous cell carcinoma (30.7%). However, in an extensive pathological examination of 47 CPFE patients with concurrent lung cancer, Girard et al. found that 38 of them (81%) had lung cancer, with 17 cases (36%) being squamous cell carcinoma and 14 cases (30%) being adenocarcinoma [29]. Additionally, Koo et al. [30] conducted a meta-analysis and reported that CPFE combined with lung cancer primarily occurs in elderly males with a history of smoking, with squamous cell carcinoma being the predominant type. The viewpoint that COPD patients are more prone to squamous cell carcinoma is supported by numerous studies. For instance, Bozinovski et al.'s analysis [31] suggests that abnormal inflammation and immune responses are common underlying factors in COPD patients' susceptibility to squamous cell carcinoma. Zhang et al. [32] confirmed through a Mendelian randomization study that airflow limitation (FEV1/ FVC<0.7) is an independent predictor for lung squamous cell carcinoma. Liu et al. [33] conducted an analysis of patients with IPF and concurrent lung cancer, revealing that 45.65% of IPF patients had adenocarcinoma.

Our research revealed that CPFE patients were more prone to develop lung cancer in the lower lobes of the lungs, particularly in areas affected by pulmonary fibrosis. This pattern accounted for 60% of cases in the CPFE+LC group. In contrast, COPD patients tended to have lung cancer occurrences predominantly in the upper lobes of the lungs. The differences in the locations of lung cancer in these two groups were statistically significant. Kwak et al.'s study [34] also supports the notion that lung cancer associated with CPFE is more likely to occur in the subpleural region, closer to dense fibrotic areas. This finding aligns with the conclusion reached by Liu et al. [33], who found that lung cancer in IPF patients primarily occurs in the peripheral and lower lobes, consistent with the affected areas of IPF. Additionally, Bae et al. [35] reported that in COPD patients, lung cancer is most likely to occur in the upper lobes of both lungs, with an odds ratio of 1.77 when compared to the lower/ middle lobes. This observation implies that emphysema may not have an additional impact on CPFE- related lung cancer and lends support to the potential relationship between cancer development and fibrotic regions. In addition to these factors, the subtypes of emphysema are also correlated with lung cancer [36].

	CPFE + LC (n = 28)	COPD + LC (n = 78)	Statistic	Р
Immune-related Adverse Events	10 (35.7%)	19 (24.4%)	χ ² =1.337	0.248
Interstitial pneumonia	5 (50.0%)	7 (36.8%)		
Pituitary insufficiency	2 (20.0%)	0		
Liver dysfunction	0	3 (15.8%)		
Renal insufficiency	1 (10.0%)	2 (10.5%)		
Diarrhea	0	2 (10.5%)		
Rash	1 (10.0%)	1 (5.3%)		
Cystitis	1 (10.0%)	0		
Myositis	0	1 (5.3%)		
Intracranial edema	0	1 (5.3%)		
Pancreatitis	0	1 (5.3%)		
Hearing loss	0	1 (5.3%)		

Table 4 Comparison of incidence and specific immune-related adverse events in CPFE+LC versus COPD+LC patients

The imaging findings of CPFE typically include upperlobe emphysema and lower-lobe interstitial fibrotic pattern. Previous studies have shown differences in the distribution of the emphysema types between CPFE and COPD. The emphysematous change in COPD is usually centrilobular, while in CPFE paraseptal emphysema is much more frequent [5, 37–39]. In our study, among CPFE+LC patients, paraseptal emphysema was the most common subtype, accounting for 41.33% of cases, while in COPD+LC patients, centrilobular emphysema was more prevalent, making up 60.99% of cases, which was in accordance with previous studies. Oikonomou et al's study [40] demonstrated that CPFE patients with paraseptal emphysema most commonly show a higher extent of fibrosis with a UIP pattern, while centrilobular emphysema may be associated with a higher extent of emphysema and an NSIP pattern, indicating a stronger association of paraseptal emphysemas with typical UIP pattern of fibrotic change. Moreover, it is worth noting that some research has found that the presence of paraseptal emphysema increases the risk of adenocarcinoma in COPD patients [41]. However, González et al. [42] discovered in a lung cancer screening project in Spain that airflow obstruction is associated with an increased risk of lung cancer, but this risk is reduced in the presence of paraseptal emphysema. This may help explain why paraseptal emphysema is less prevalent in COPD+LC patients.

For patients in the CPFE+LC group, regardless of whether they received chemotherapy alone or a combination of chemotherapy and immunotherapy, the median PFS of first-line treatment was 6.0 months. Our research reveals that lung cancer patients with CPFE may not derive additional benefits from immunotherapy. Tan et al. [43] reported two cases of lung cancer patients with CPFE, both of whom received a treatment of chemotherapy combined with immunotherapy. After treatment, both patients experienced a significant reduction in tumor size. However, one eventually died from worsening acute interstitial lung disease caused by immunotherapy, while the other died due to tumor infiltration after discontinuing immunotherapy. Our study found that 35.7% of the CPFE+LC group receiving immunotherapy experienced immune-related AEs, while the COPD+LC group accounted for 24.4%. There was no significant difference in the incidence of immune-related AEs in two groups, indicating similar tolerability to immunotherapy in both groups. We also found that both the CPFE+LC group and the COPD+LC group had interstitial pneumonia as the predominant immune-related AE. Immune checkpoint inhibitors can lead to immune-related AEs, with interstitial lung disease being one of the more severe adverse events among them [44].

In our study, we observed that the CPFE+LC group exhibited significantly higher levels of tumor markers such as CEA and NSE compared to those in the COPD+LC group. These elevated levels of tumor markers may suggest a greater tumor burden or more extensive tumor invasion. Furthermore, in accordance with the research by Koo et al. [30], lung cancer in CPFE patients is often diagnosed at advanced stages, possibly due to fibrosis and emphysema masking the symptoms of the tumor, resulting in misdiagnosis or delayed diagnosis. This not only exacerbates the already compromised lung function but also increases the complexity of surgical treatment. In addition, numerous studies have identified common signaling pathways in lung cancer and pulmonary fibrosis. For example, connexin 43 has been found to exhibit reduced expression or expression loss in both conditions [45]. Additionally, molecules involved in the regulation of the Wnt/ beta-catenin signaling pathway are overexpressed in the lung tissues of patients with lung cancer and pulmonary fibrosis, contributing to processes such as lung remodeling and carcinogenesis [46].

We found that in advanced cancer patients with TNM staging ranging from IIIB to IV, the CPFE+LC group receiving first-line treatment had a significantly shorter median PFS of 6.0 months compared to 9.0 months in the COPD+LC group. This trend of poorer prognosis was consistently observed across different histological types: patients with NSCLC who had combined CPFE exhibited worse outcomes than those with combined COPD, and a similar prognostic pattern was observed in patients with SCLC. These results further emphasize the greater challenges faced by the CPFE+LC group in terms of treatment response and disease progression. Existing research indicates that CPFE+LC patients have a significantly shorter median survival compared to patients with lung cancer alone or lung cancer with emphysema [28]. Kumagai et al. [47] further revealed that CPFE patients exhibit lower tolerance to tumor chemotherapy, with a higher recurrence rate and markedly shorter Overall Survival (OS) in NSCLC.

Our study has several limitations. First, as a singlecenter, retrospective analysis, the generalizability of our findings may be limited and subject to selection bias. Second, we excluded patients who underwent surgical treatments, some of whom received neoadjuvant or adjuvant chemotherapy due to their earlier TNM stages, potentially skewing treatment efficacy results. Furthermore, the brief study period and small sample size may have introduced bias. For example, the fact that all patients in the CPFE+LC group were male could indicate a selection bias. Also, the lower incidence of CPFE compared to COPD resulted in a small number of lung cancer patients with CPFE, leading to a numerical imbalance that might bias the results. Future studies should expand the sample size and include detailed survival analyses for each histological type of lung cancer. Finally, the short followup duration limited our ability to assess overall survival, restricting our analysis to PFS within the available followup period.

Conclusion

In conclusion, this retrospective study underscores the distinct clinical, pathological, and functional characteristics of lung cancer patients with CPFE or COPD. Notable differences were observed in tumor pathology, pulmonary function parameters, and treatment responses between the CPFE+LC and COPD+LC groups. These findings highlight the necessity for tailored diagnostic and therapeutic approaches in managing lung cancer within these patient populations. While our study provides valuable insights, its retrospective, single-center nature suggests the need for further multicenter, prospective research to validate and expand upon these findings. Ultimately, this study emphasizes the critical need for heightened clinical awareness and individualized treatment strategies for lung cancer in the context of CPFE or COPD.

Abbreviations

LC	Lung cance
LC	Lung canc

CPFE	Combined pulmonary fibrosis and emphysema
COPD	Chronic obstructive pulmonary disease
PFS	Progression-free survival
NSCLC	Non-small cell lung cancer
SCLC	Small cell lung cancer
IPF	Idiopathic pulmonary fibrosis
HRCT	High-resolution computed tomography
ECOG	Eastern Cooperative Oncology Group
AEs	Adverse reactions
HR	Hazard ratio
OS	Overall survival

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Author contributions

Yuying Wei: Conceptualization, Data curation, Formal analysis, Methodology, Resources, Software, Validation, Visualization, original draft, Writing - review & editing. Liuqing Yang: Data curation, Investigation, Project administration, Resources, Writing - review & editing. Qing Wang: Conceptualization, Methodology, Supervision, Validation, Writing - review & editing. All authors read and approved the final manuscript.

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Data availability

The datasets generated and/or analyzed during the current study are not publicly available but are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

The Ethics Board of the First Affiliated Hospital of Zhejiang University approved this study. Written informed consent was waived by the Ethics Board of the First Affiliated Hospital of Zhejiang University due to the retrospective nature of the study.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

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